# *Cutibacterium acnes*: A Proposal for Veterinary Medicine

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Abstract:- *Cutibacterium acnes* (*C. acnes*) is, by far, a highlighted immunomodulator that in recent years has gained prominence for its performance in experimental clinical studies, prophylaxis, and therapeutics of infectious diseases and neoplasms.

If, on the one hand, it can act as an opportunistic pathogen, on the other, can be used as an and oncological immunomodulator the in area. Furthermore, it is also considered a proposal to strengthen the immune system to combat severe acute respiratory syndrome caused by coronavirus 2 (Sars-Cov-2). In other words, its application extends from bacteria, fungi, and parasitic protozoa to viruses. However, even with its growing applications in human medicine, little has been reported in the field of veterinary medicine.

With that said, this review has the purpose of directing the scientific community to the potential of *C*. *acnes* aimed attreating pathology, especially in animals.

*Keywords:- Cutibacterium Acnes, Immunotherapy, Animals, Veterinary Medicine.* 

# I. INTRODUCTION

In 1974, the synonymy between *Corynebacterium* parvum (*C. parvum*) and *Propionibacterium acnes* (*P. acnes*) was established [1]. Later, in 2016 [2], with comparative studies of DNA homology of bacterial genera, *P. acnes* was renamed *Cuticubacterium acnes* [3], to differentiate it from other species of the genus *Propionibacterium* [4]. Before the widespread use of *C. acnes*, *Bacillus Calmette Guerin* (BCG) was used as an immunotherapy to treat neoplasms, but due to the systemic risks of using BCG, researchers found in *C. acnes* a safer alternative without systemic risks but with great similarity [5-7].

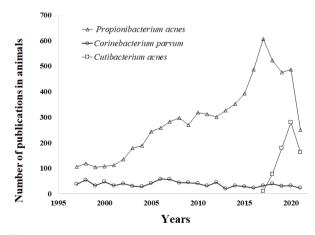
Considering its peculiarities, *C. acnes* is a commensal bacterium, facultative anaerobic [8] whose final fermentation product includes propionic acid. Being gram-positive, it is present in the human skin microbiota (sebaceous follicle) [9-11] and can trigger some pathologies, such as inflammatory dermatoses, when that goes out of balance [12-15]. It can modulate various functions of the innate and adaptive immune response, indulging the reduction of systemic infections from multiple origins such as viral 16, bacterial, fungal, or protozoan [17,18], by preventing the evolution of the virulence process or

the infective process. Owing to its role in promoting immune defenses, it is currently suggested in the treatment of acute respiratory syndrome, for cases of Covid-19 [19], as a stimulant of the immune mechanism [20], immunomodulator [21], treatment of warts [22], treatment of neoplasms [23], among other pathologies. Besides these encouraging results, there are still punctual studies, not so comprehensive, on the relevance of the use of *C. acnes* for therapeutic purposes used in veterinary medicine, one of the primaries focuses addressed in this review.

# II. CUTIBACTERIUM ACNES

#### Experimental studies in animals

Few scientific papers are available for consultation when it comes to therapeutic actions using *C. acnes* applied to animals. A search on the *Science Direct* platform (https://www.sciencedirect.com/) showed the number of publications on *C. acnes* aimed at its application in veterinary medicine, using the keywords: *Corynebacterium parvum*, *Propionibacterium acnes* and *Cutibacterium acnes* in the period between 1997 and the first half of 2021 (*May*), (**Figure 1**).



**Fig. 1**. Evaluation of scientific publications presented by *Science Direct* for publications with *C. acnes* applied to animals, (a search carried out in May/2021).

The immunogenic potential using *C. acnes* lysate started to be investigated in clinical studies in rats [24], showing a relationship between the activation of the immune system and the stabilization, regression or increase in the longevity of animals transplanted with a high-rate tumor cells mortality. This was confirmed by Andre; Zitvogel; Sabourin, [25], where the authors studied the performance of two vaccines

(*Wellcome Reagents*, Ltd. and *Institut Merieux* - Lyon, France), containing a suspension of inactivated *C. acnes*, to indulge immune reactivity in mice with breast carcinoma. The animals were treated for a period of 3 to 12 days with subcutaneous applications of bacterial lysate, by intravenous, intraperitoneal and intratumoral administrations. Tumor regression and inactivation were observed. The authors report that intratumoral injection thirteen days before surgical removal prolongs the survival or cure of treated animals.

Braga and collaborators [26], treated subcutaneoussly (murine model) mice sensitized by ovalbumin, inducing so an allergic pulmonary response. Therapy followed the application of 700 mg of C. acnes, resuspended in 350 ml of saline, once a week (subcutaneously), for a period of three weeks. In the second week, the animals were treated for seven days with a suspension of C. acnes, and in the last week, the animals were sensitized with ovalbumin. It was observed that C. acnes acted to modulate the late phase-type I hypersensitivity reaction. It is reported that subcutaneous injections of C. acnes lysate over three weeks induced specific antibody responses, and increased the phagocytic capacity of peritoneal macrophages. In a period, shorter than 24 h, after a single intraperitoneal inoculation, there was a significant increase in macrophages, immature dendritic cells, and Natural Killer (NK) cell population. Prompt treatment before sensitization modulated the specific immune response, promoting a Th1 response, preventing hypersensitivity to the allergen.

Abreu and collaborators [27], presented the case study where dogs affected with oral papillomatosis were treated with an association between Vincristine sulfate and suspension of C. acnes. Also considering the lack of an effective treatment for this purpose, the authors used C. acnes as an immunostimulant, together with liposaccharides from Escherichia coli. The therapeutic protocol was performed by weekly applications of Vincristine, intravenously, starting with the first dose of 0.025 mg/kg; in the second week, 0.035 mg/kg and thereafter, 0.05 mg/kg. 1.0 ml of the suspension of C. acnes was applied intramuscularly (1.0 ml per kg) every 15 days. It was observed that from the third week onwards there was regression of the papillomas followed by healing after three months. Due to the success of the therapy, the authors suggest that this protocol becomes an alternative for cases resistant to the usual therapy to Thuya occidentalis.

Manickam and collaborators [28], used inactivated *C.* acnes, as an adjuvant immunizing agent in a group of calves for production of non-specific resistance to protozoan *Theileria annulata* present in ticks. Crossbred calves (*Bos indicux*  $\times$  *Bos taurus*) received 200 mg of *C.* acnes reconstituted in 2.5 ml of distilled water subcutaneously. After a period of 45, 60 or 90 days past immunization, the calves were challenged with ticks infected with *Theileria annulata*, where only mild reactions were observed. All other animals in the control group that did not receive the bacterial lysate died due to *Teileriosis* [28].

On the other hand, the authors Dinsmore and collaborators [29], reported low efficiency of C. acnes for dairy cows affected by Staphylococcus aureus, which causes infection of the cows mammary quarters. Milk obtained from milking was analyzed for a group of 51 lactating cows. Twenty-four cows received 1.0 ml injections of the C. acne suspension, twice a week, for a period of one month, while the others did not receive the bacterial suspension. 16.7% of treated cows were cured of S. aureus and 11.1% of untreated cows were cured, regardless of the use of bacterial lysate. Therefore, showing a not-so-significant effect of the immunomodulator. The authors, however, do not report the dosage applied in terms of bacterial lysate mass. The application and efficiency of Cutibacterium acnes were investigated experimentally in animals and are shown in Table 1.

## Case Study

In the process of a bibliographic survey for the construction of this review proposal, few presentations regarding "Case Studies" are available for consultation in the field of veterinary medicine. However, an interesting study presented by Abreu and collaborators [27], successfully investigated the use of *C. acnes*, as an adjuvant in the treatment of Papillomatosis, as images are shown in Figure 2. The authors' used therapy consisted of the use of Vincristine associated with an immunostimulant based on C. acnes lysate and Escherichia coli liposaccharides. The bacterial lysate was administered intramuscularly every 15 days, at a dosage of 1.0 ml (1.0 ml/10 kg), for a period of six months, after intravenous administration of Vincristine sulfate at a dosage of 0.025 mg/kg in the first week; 0.035 mg/kg in the second week; and subsequently 0.05 mg/kg. A full recovery of the animals was reported.



Fig. 2. Results obtained by Abreu and collaborators [27], using *C. acnes* as an adjuvant in the treatment of canine oral *papillomatosis*. (1-4) Progressive evolution over 6 months with 15/15 days of administration of *C. acnes* with adjuvant/ Vincristine.

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Therapy	Application
Adjuvant/	Primary tumors (mouses) <sup>(30)</sup>
Immunostimulants	
Nano-immunotherapy	Melanoma (mouses) <sup>(31)</sup>
Nano-immunotherapy	Murine hepatitis (mices) <sup>(32)</sup>
Immunotherapy	Oral melanoma (dogs) <sup>(33)</sup>
Immunotherapy	Dermatitis/Pyoderma (dogs) <sup>(34)</sup> Tumor (mices) <sup>(35)</sup>
Immunotherapy	Tumor (mices) <sup>(35)</sup>
Application under bone marrow-derived dendritic cells	Tumor (mices) <sup>(36)</sup>
Immunomodulator/Protector against infections	Rabies virus infection (mices) (37)(38)
Fragmentation of C. acnes	Taxological risk assessment
by nanotechnology	(monkeys) <sup>(39)</sup>
Anti-inflammatory	Sepsis / Inflammation (mices) (40)
Immunotherapy	Inhibition of tumor (mouses) (41)
Protective effect	Hepatitis (mouses) <sup>(42)</sup>
Immunomodulator /viral	Infection with Herpesvirus
resistance	hominis (mices) <sup>(43)</sup>
Adjuvant with vaccine	Fungo Paracoccidioidomycosis
,	(mouses) <sup>(44)</sup>
Adjuvant with vaccine	Leishmania major <sup>(45)</sup>
Immunotherapy	Malignant melanoma (dogs) <sup>(46)</sup>
Immunotherapeutic agent	Inhibition of <i>Sarcoma</i> <sup>(47)</sup>
Antitumor, Anti- Angiogenesis and Immunomodulatory Effects	Breast cancer (mices) (48)
Anti-tumor/Immunological adjuvant	Tumor (mices) <sup>(49)</sup>
Stimulant of the reticuloendothelial system	Burn treatment (mices) <sup>(50)</sup>
Infection prophylactic	Pleuropneumonia (porcines) <sup>(51)</sup>
Adjuvant/	Inflation reduction (mices) <sup>(52)</sup>
Immunomodulator	
Pre-treatment / antitumor	Solid tumor reduction (mices) <sup>(53)</sup>
Immunomodulator	Brucellosis (guinea pigs) <sup>(54)</sup>
Immunomodulator	Papillomatosis (dogs) <sup>(55)(56)</sup>

Table 1. Experimental s	studies using	<i>Cutibacterium acnes</i> in	
animals.			

#### III. ACTION MECHANISM IN TUMORS

Over decades, studies with *C. acnes* have shown its antitumor action, demonstrating the inhibitory or regressive activity, under conditions of neoplasms, in animals and humans [57-63], but the exact mechanism of action of *C. acnes* in inducing tumor growth delay or regression is still unknown. However, its immunomodulatory effects are reported, mainly due to its ability to activate cellular functions of the

#### Immune Response

When *C. acnes* lysate is injected into the body, the immune system "perceives" the presence of these particles, initially, through phagocytic activity and possible cell types: macrophages, neutrophils, dendritic cells, leukocytes (NK), mast cells, basophils, eosinophils or keratocytes [70]. C. acnes simulates a pathogen that crosses the skin barrier, which activates innate immunity, using one of the cell types phagocytosis, PPR (Pattern After mentioned above. Recognition Receptors), TLR (Toll-Like), NOD (Domain Nucleotide Oligomerization), RIG-I (Retinoic Acid Inducible Gene) and C-Type Lecithin receptors are expressed [71]. PPR recognizes different pathogens that are associated with the PAMP pattern (Pathogen-Associated Molecular Pattern) or cause cell damage (DAMP) and consequently induce the production of pro-inflammatory substances (cytokines, TNFalpha and IFN-gamma and chemokines (which recruit more phagocytic cells).

The starting point for immunostimulation from C. acnes lysate occurs when NK cells are activated and this is the main proposal for its use. At this moment, a quick response is stimulated to the possible bacterial, fungal or protozoal infection, and if it is fast enough, it will prevent the invading organism from reaching other tissues. When infection occurs by a virus, the initial immune reaction is triggered and activates the adaptive system as follows: When NK cells phagocytose the C. acne lysate, they maximize the production of initial INF'a increasing the antiviral status [72] which, concomitantly, stimulates promyelocytic leukemia nuclear bodies to associate with histone chaperones to capture the viral genome and block its replication [73-74]. Keratocytes also release IL-1 alpha and IL-36 to enhance the antiviral state, acting as early warning signals for leukocyte chemotaxis and increased cellular sensitivity to type I IFN signaling [75]. In practice, both an innate and an adaptive response are initiated concurrently, regardless of the type of invading organism.

#### Side and Adverse Effects

The use of *C. acnes* lysate in most experimental studies shows few or rare adverse reactions, thereby, its use is well tolerated. Among the reported side effects there are fever and chills [76]. Fever is an expected side effect as substantial amounts of liposaccharides are being injected and the chills are an effect of the fever. However, some authors have observed adverse reactions in mice treated with the bacterial lysate, such as the development of intrahepatic granulomas and scleromas in injected areas [77].

The formation of granulomas and scleromas can be related to applications of exceedingly high concentrations in the same place, which causes an intense inflammatory process in the region of application. It should be noted, however, the dosage to be applied in each specific case, especially regarding its

combination with other substances, such as cyclophosphamide, for example [78]. In an attempt to minimize possible adverse reactions, studies in the field of nanotechnology have been developed based on the encapsulation of bacterial lysate, with the advantage of being easily absorbed and released in low concentrations in the body [31].

# Clinical Application

C. acne lysates were used in peritoneal applications [79], intravenously or directly infiltrated into the neoplastic nucleus, in animals [80] and humans [60]. As for their use on a global scale, C. acnes lysates have never been industrially traded for human use outside Brazil, where the LAFEPE laboratory (Pharmaceutical Laboratory of Pernambuco) had the registration of C. acnes lysate with the trade name of IMUNOPARVUM®, its orientation for use is 5 deep subcutaneous applications with volumes from 0.5 ml to 1.0 ml for 7/7 days. After this period, an assessment of cellular immunity with PPD (Tuberculin or Mantoux reaction). Candidine, or Tricophytin tests must be performed, if it reaches normal levels of cutaneous reactivity. The sequence of applications must be carried out for 15/15 days, being the duration defined by medical criteria according to the package insert. Another product, in this case only for veterinary use, INFERVAC® has also been marketed. In this case, indicated doses were 2.0 ml for animals weighing up to 10 kg and at intervals of 48 hours, the concentration of C. acnes doses reaches 400 mg / ml [81], it is no longer marketed.

Currently (2021), in Brazil, only the *Laboratory of Allergenic Extracts* LTDA, holds the registration of the *C. acnes* lysate, with the name PARVULAN® at the concentration 2.0 mg/ml in 5.0 ml vials and application guidance of 0.5 ml for 7/7 days (ten doses). In the country, most of the *C. acnes* lysate is marketed in a manipulated way and used as an adjuvant in immunotherapy formulations. A major difference between the use of bacterial lysates from the upper respiratory tract and *C. acnes* lysate is that the latter is applied in an injectable way, specifically deep subcutaneously, causing a systemic action, contrary to bacterial lysates.

# IV. CONCLUDING REMARKS

The *C. acnes* lysate shows immense potential in different applications, from solving an immunological problem of "low immunity", improving the body's defense capacity against bacteria, fungi, viruses and protozoa, to an effective application in different grades of neoplasms. Its activity seems to be dose and interval-dependent, behaving as an immunomodulator at low doses and long application intervals (interesting use to control chronic inflammatory processes) and as an immunostimulator with high doses and short intervals. This allows for appropriate management for different pathologies and can even be used as an adjuvant, enhancing the synergism of vaccines and many medications (antibiotics, antivirals). Its use in neoplasms is better in closer or intratumoral doses, that is, it is limited to neoplasms that are within reach of the syringe needle. Application in veterinary medicine is very promising, as it is much less invasive than surgery and does not leave scars, and reduces the chances of metastases, in addition to the fact that surgical anesthetics often work as immunosuppressants, which is bad for recovery, it can still be monthly, quarterly or biannual doses with prophylactic function.

# Conflicts of interest

There are no conflicts of interest associated with the publication of this article.

## REFERENCES

- Cummins C. S. Johnson JL, "Corynebacterium parvum: a Synonym for Propionibacterium acnes?" Journal Gen Microbiol. 1972, 8 (972): pp. 433–442.
- [2]. Platsidaki E. Dessinioti C, "Recent advances in understanding Propionibacterium acnes (Cutibacterium acnes) in acne". F1000Research. 2018,7, pp. 1-12.
- [3]. Dréno B. Pécastaings S. Corvec S. Veraldi S. Khammari A. Roques. C, "*Cutibacterium acnes (Propionibacterium acnes) and acne vulgaris: a brief look at the latest updates*". J Eur Acad Dermatology Venereol. 2018, 32, pp. 5-14.
- [4]. Castillo DE. Nanda S. Keri JE. "Propionibacterium (Cutibacterium) acnes Bacteriophage Therapy in Acne: Current Evidence and Future Perspectives". Dermatol Ther (Heidelb). 2019, 9(1): pp. 19–31.
- [5]. Megid, J. Peraçoli; MTS, Curi; RR, Zanetti; CR, Cabrera; WH, R.Vassao; F.H I. "Effect of bacillus of calmette-Guérin, Avridine and Propionibacterium acnes as immunomodulators on Rabies in mice". Rev Inst Med trop S Paulo. 2012, 41(2): pp.107–114.
- [6]. Sher NA, Chaparas SD, Greenberg LE, Bernard S. "Effects of BCG, Corynebacterium parvum, and methanol extraction residue in the reduction of mortality from Staphylococcus aureus and Candida albicans infections in immunosuppressed mice". Infect Immun. 1975, 12(6): pp. 1325–30.
- [7]. Lopes CAC SC. "New descriptions on the use of Corynebacterium parvum in patients with infectious diseases". Ministry of Education and Culture, Federal University of Pernambuco. 1984, (1): pp. 1–15.
- [8]. Alnabati NA, Al-Hejin AM, Noor SO, Ahmed MMM, Abu-Zeid M, Mleeh NT. "The antibacterial activity of four Saudi medicinal plants against clinical isolates of Propionibacterium acnes". Biotechnol Biotechnol Equip. 2021, 35(1): pp. 415–24.
- [9]. Yang J, Tsukimi T, Yoshikawa M, Suzuki K, Takeda T, Tomita M, et al. "Genotyping of Microbial Samples from Possessions Contributes to Owner Identification". Hostmicrobe Biol. 2019, 4(6): pp. 1–15.
- [10]. Grange PA, Raingeaud J, Morelle W, Marcelin AG, Calvez V, Dupin N. "Characterization of a Propionibacterium acnes Surface Protein as a Fibrinogen-Binding Protein". Sci Rep. 2017, 7(1): pp. 1–14.
- [11]. Davidsson S, Mölling P, Rider JR, Unemo M, Karlsson MG, Carlsson J, et al. "Frequency and typing of Propionibacterium acnes in prostate tissue obtained

from men with and without prostate cancer". Infect Agent Cancer. 2016, 11(1): pp. 1–10.

- [12]. Okazaki F, Wakiguchi H, Korenaga Y, Nakamura T, Yasudo H, Uchi S, et al. "A novel mutation in earlyonset sarcoidosis/Blau syndrome: an association with Propionibacterium acnes". Pediatr Rheumatol. 2021,19(1): pp. 1–9.
- [13]. Beijer E, Seldenrijk K, Eishi Y, Uchida K, Damen J, Grutters JC, et al. "Presence of Propionibacterium acnes in granulomas associates with a chronic disease course in Dutch sarcoidosis patients". ERJ Open Res. 2021, 7(1): pp. 1–10.
- [14]. Jha SC, Sairyo K. "The role of propionibacterium acnes in and modic type 1 changes: A literature review". J Med Investig. 2020, 67, pp. 21–26.
- [15]. Fassi Fehri L, Mak TN, Laube B, Brinkmann V, Ogilvie LA, Mollenkopf H, et al. "Prevalence of Propionibacterium acnes in diseased prostates and its inflammatory and transforming activity on prostate epithelial cells". Int J Med Microbiol. 2011, 301(1): pp. 69–78.
- [16]. Palmieri B, Vadalà M, Roncati L, Garelli A, Scandone F, Bondi M, et al. "The long-standing history of Corynebacterium parvum, immunity, and viruses". J Med Virol. 2020, 92(11): pp. 2429–39.
- [17]. E. Ghadirian; P. A. L. Kongshavn. "Protection of mice against intestinal amoebiasis with BCG, corynebacterium parvum and Listeria monocytogenes". Parasite Immunol. 1986, 8, pp. 663–7.
- [18]. Korbelik M, Hode T, Lam SSK, Chen WR. "Novel Immune Stimulant Amplifies Direct Tumoricidal Effect of Cancer Ablation Therapies and Their Systemic Antitumor Immune Efficacy". Cells. 2021, 10(3): p. 492.
- [19]. Roncati L, Vadalà M, Corazzari V, Palmieri B. "COVID-19 vaccine and boosted immunity: nothing ad interim to do?" J Pre-proofs. 2020, 38(48): pp. 7581–7584.
- [20]. Palmieri B, Bondi M, Cermelli C, Specialties S, Emilia R, Clinic S, et al. "Propionibacterium acnes: A putative immunemodulating weapon against the coronavirus impending epidemy". Int J Innov Sci Res Technol. 2020, 5(2): pp. 431–40.
- [21]. Hokland P, Ellegaard J, Heron I. "Immunomodulation by Corynebacterium parvum in normal humans". J Immunol. 1980, 124(5): pp. 2180-2185.
- [22]. Nasser N. "Treatment of common warts with the immune stimulant Propionium bacterium parvum". An Bras Dermatol. 2012, 87(4): pp. 585-589.
- [23]. Knapp RC, Berkowitz RS. "Corynebacterium parvum as an immunotherapeutic agent in ovarian cancer model". Am J Obstet Gynecol. 1977, 128(7): pp. 782-786.
- [24]. Dagnelie MA, Khammari A, Dréno B, Corvec S. "Assessment of seven protocols to prepare Cutibacterium acnes bacterial lysates to measure its immunogenic potential and review of the literature". Anaerobe. 2019, 57, pp. 75–81.
- [25]. Andre F, Zitvogel L, Sabourin J-C. "Treatment of cancer using Corynebacterium parvum: Similarity of two prepations in four animal tumor models". Am cancer Soc. 2008, 46(1): pp. 685–91.

- [26]. Braga EG, Ananias RZ, Mussalem JS, Squaiella CC, Longhini ALF, Mariano M, et al. "Treatment with Propionibacterium acnes modulates the late phase reaction of immediate hypersensitivity in mice". Immunol Lett. 2003, 88(2): pp. 163–9.
- [27]. Abreu CB De, Eduardo L, Oliveira D De, Baeta P, Cerqueira JA, Schulien T, et al. "Association of vincristine sulfate and Propionibacterium acnes in the treatment of canine oral papillomatosis" – Case report. 2015, (21). p. 11–7.
- [28]. Manickam R, Dhar S, R.P. Singh. "Non-specific immunization against bovine tropical theileriosis (Theileria annulata) using killed Corynebacterium parvum". Vet Parasitol. 1983, 13(2): pp. 115–9.
- [29]. Dinsmore R.P., Cattell M.B., Stevens R.D., Gabel C.S., Salman M.D., and Collins J.K. "Eficacy of a Propionibacterium Acns Immunostimulant for Treatment of Chronic Staphylococcus Aureus Mastitis." Journal of Dairy Science. 1995. 78(9): pp. 1932–36.
- [30]. Miao Wang, Jie Rao, Meng Wang, Xiaosong Li, Kaili Liu, Mark F. Naylor, Robert E. Nordquist, Wei R. Chen and FZ. "*Cancer photo-immunotherapy: from bench to bedside. Theranostics*". 2021, 11(5): pp. 2218–31.
- [31]. Gao S, Liu C, Qu S, Song J, Li J, Zhang P, et al. "Noncell Corynebacterium parvum generated by nanotechnology: A promising immunomodulator with less side effects". Int Immunopharmacol. 2007,7(10): pp. 1334–42.
- [32]. Liu C, Zhang L, Gao S, Qu Z, Wang Q, Zhu F, et al. " NCPP treatment alleviates ConA-induced hepatitis via reducing CD4+T activation and NO production". Immunopharmacol Immunotoxicol. 2012, 34(6): pp. 962-967.
- [33]. Macewen EG, Patnaik AK, Harvey HJ, Hayes AA, Matus R. "Canine oral melanoma: Comparison of surgery versus surgery plus Corynebacterium parvum. Cancer Invest. 1986, 4(5): pp. 397–402.
- [34]. A. M. Becker, T. A. Janik, DVM, E.K Smith, C. A. Sousa and BAP. "*Dematite canina C. Parvum*". J Vet. 1989, 3(1): pp. 26–30.
- [35]. Wilson RG. "Simple estimation of macrophage activity in tumour-bearing animals treated with Corynebacterium parvum". Br J Exp Pathol. 1981, 62, pp. 347–9.
- [36]. Liu C, Gao S, Qu Z, Guo C, Wu P, Shi Y, et al. "Effect of non-cell corynebacterium parvum on differentiation and maturation of bone marrow-derived dendritic cells". Immunol Invest. 2012, 41(8): pp. 820–30.
- [37]. Megid J, Kaneno R. "Natural killer activity in mice infected with rabies virus and submitted to P. acnes (Propionibacterium acnes) as immunomodulator". Comp Immunol Microbiol Infect Dis. 2000, 23(2): pp. 91–7.
- [38]. Megid J, Cremonini DN, Leomil H. "Distribution of rabies virus in infected mice, vaccinated and sumitted to *P. acnes as immunomodulator*". Comp Immunol Microbiol Infect Dis. 2002, 25(4): pp. 237–48.
- [39]. Li BQ, Dong X, Fang SH, Yang GQ, Gao JY, Zhang JX, et al. "Systemic toxicity of non-cell corynebacterium parvum (CP) in monkeys". J. Toxicol Sci. 2010, 35(3): pp. 279–86.

- [40]. Silva JBNF, de Oliveira SKM, Campos IA, de Carvalho-Júnior CHR, Coutinho T da C, Silva TG. "Propionibacterium acnes-killed attenuates the inflammatory response and protects mice from sepsis by modulating inflammatory factors". Brazilian J Infect Dis. 2013, 17(1): pp. 20–26.
- [41]. Halpern BN, Biozzi G, Stiffel C, Mouton D. "Inhibition of Tumour Growth by Administration of Killed Corynebacterium parvum". Nature. 1966, 212: pp. 853– 854.
- [42]. Schindler L, Streissle G, Kirchner H. "Protection of mice against mouse hepatitis virus by Corynebacterium parvum". Infect Immun. 1981, 32(3): pp. 1128–1131.
- [43]. Glasgow LA, Fishbach J, Bryant SM, Kern ER. "Immunomodulation of host resistance to experimental viral infections in mice: Effects of Corynebacterium acnes, Corynebacterium parvum, and bacille Calmette Guerin". J Infect Dis. 1977, 135(5): pp. 763–770.
- [44]. Ea M, Ja DCO, Em M, Maf C, Rap N, Rc R, et al. "Efficacy of Chitosan as Vaccine Adjuvant Against Paracoccidioidomycosis in Efficacy of Chitosan as Vaccine Adjuvant Against Paracoccidioidomycosis in Mice". J Immunol Infect Dis. 2018, 5(5): pp. 1–16.
- [45]. Yang DM, Rogers M V., Liew FY. "Identification and characterization of host-protective T-cell epitopes of a major surface glycoprotein (gp63) from Leishmania major". Immunology. 1991, 72(1): pp. 3–9.
- [46]. Atherton MJ, Morris JS, McDermott MR, Lichty BD. "Cancer immunology and canine malignant melanoma: A comparative review". Vet Immunol Immunopathol. 2016,169, pp. 15–26.
- [47]. Cham BE. "Combination Treatment with BEC and Cisplatin Synergistically Augments Anticancer Activity and Results in Increased Absolute Survival". J Cancer Ther. 2020, 11(08): pp. 470–482.
- [48]. Talib WH, Saleh S. "Propionibacterium acnes augments antitumor, anti-angiogenesis and immunomodulatory effects of melatonin on breast cancer implanted in mice". PLoSOne. 2015, 10(4): pp. 1-13.
- [49]. Turney TH, Harmsen AG, Jarpe MA. "Modification of the antitumor action of Corynebacterium parvum by stress". Physiol Behav. 1986, 37(4): pp. 555–558.
- [50]. Kirov A, Hahn H, Müller F, Koslowski L, Roth C, Schmidt K. "Therapeutic effects of Corynebacterium parvum in experimental burn disease of mice". Burns. 1979, 6(1): pp. 45–47.
- [51]. Liu J, Ma Q, Yang F, Zhu R, Gu J, Sun C, et al. "Veterinary Microbiology". 2017, 205(4): pp. 14–21.
- [52]. Gambero M, Teixeira D, Butin L, Ishimura ME, Mariano M, Popi AF, et al. "Propionibacterium acnes induces an adjuvant effect in B-1 cells and affects their phagocyte differentiation via a TLR2-mediated mechanism". Immunobiology. 2016, 221(9): pp. 1001–1011.
- [53]. Castro JE. "Antitumour effects of Corynebacterium parvum in mice". Eur J Cancer. 1974,10(2): pp: 121–127.
- [54]. With I, "Propionibacterium K, In A. Immunomodulation with killed in guinea pigs simultaneously vaccinated with strain 19". Vet Immunol Immunopathol. 1986, 13(9): pp. 71-84.

- [55]. Megid J, Dias JG, Aguiar DM, Nardi G, Silva WB, Ribeiro MG. "*Treatment of canine papillomatosis with Propionibacterium acnes*". Arq Bras Med Vet e Zootec. 2001, 53(5): pp. 574–576.
- [56]. Abreu CB De, Eduardo L, Oliveira D De, Baeta P, Cerqueira JA, Schulien T, et al. "Associação de sulfato de vincristina e Propionibacterium acnes no tratamento de papilomatose oral canina" – Relato de Caso. "Association of vincristine sulfate and Propionibacterium acnes in the treatment of canine oral papillomatosis - case report". Vet Not. 2015, 21(1): pp. 11–7.
- [57]. E S Dye, R J North CDM. "Mechanisms of anti-tumor action of Corynebacterium parvum. I. "Potentiated tumor-specific immunity and its therapeutic limitations". J Exp Med. 1981, 154(9): pp. 609–620.
- [58]. Karashima A, Taniguchi K, Yoshikai Y, Nomoto K. "Alteration in Natural Defense Activity Against NKSusceptible B16 Melanoma Cells after Treatment with Corynebacterium parvum". Immunobiology. 1991, 182(5): pp. 414–24.
- [59]. Millman I, Scott AW, Halbherr T. "Antitumor Activity of Propionibacterium acnes (Corynebacterium parvum) and Isolated Cytoplasmic Fractions". Cancer Res. 1977, 37(11): pp. 4150–4155.
- [60]. Acute IN, By ADCC. "Immunomodulation of NK and ADCC by Corynebacterium parvum in acute myeloid leukaemia patients". Leuk Res. 1985, 9(1): pp. 175–184.
- [61]. Fox M, Ph D, Woods RL, Tattersall MHN. "A Randomized study of adjuvant immunotherapy with levamisole and corynebacterium parvum in operable non-small cell lung cancer". Immunother Immunol. 1980, 6, pp. 1043–1045.
- [62]. Murahata RI, Zighelboim J. "Systemic administration of Corynebacterium parvum during sensitization to tumor alloantigen-modified response to rechallenge". Cancer Immunol Immunother. 1980, 8(4): pp. 257–261.
- [63]. Karrer K, Rella W, Goldin A. "Surgery plus Corynebacterium parvum immunotherapy for Lewis lung carcinoma in mice". Eur J Cancer. 1979, 15(6): pp. 867– 873.
- [64]. Silveira G da P, Ishimura ME, Teixeira D, Galindo LT, Sardinha AA, Porcionatto M, et al. "Improvement of mesenchymal stem cell immunomodulatory properties by heat-killed Propionibacterium acnes via TLR2". Front Mol Neurosci. 2019, 11(1): pp. 1–14.
- [65]. Rossol S, Voth R, Brunner S, Muller WEG, Biittnera M, Gallatin H, et al. "Human peripheral blood mononuclear cells and monocytes in vitro". Eur J Immunol. 1990, 20: pp. 1761–5.
- [66]. Cooney BDR, Lewis AD, Waz W, Khan AR, Karp MP. "The effect of the immunomodulator corynebacterium parvum on hemisplenectomized mice. J Pediatr Surg. 1984,19(6).
- [67]. Fischer N, Mak TN, Shinohara DB, Sfanos KS, Meyer TF, Brüggemann H. "Deciphering the intracellular fate of Propionibacterium acnes in macrophages". Biomed Res Int. 2013, 2013, p. 1-11.
- [68]. Chapes SK, Haskill S. "Evidence for granulocytemediated macrophage activation after C. parvum immunization". Cell Immunol. 1983, 75(2):367–77.

- [69]. Keller R, Keist R. "Induction, maintenance, and reinduction of tumoricidal activity in bone-marrowderived mononuclear phagocytes by macrophageactivating lymphokines". Cell Immunol. 1986, 101(2): pp. 659–566.
- [70]. Cruvinel WDM, Júnior DM, Antônio J, Araújo P, Tieko T, Catelan T. "Immune system – Part I Fundamentals of innate immunity with". Bras J Rheumatol. 2010, 55(11): pp. 434–461.
- [71]. Bonam SR, Partidos CD, Halmuthur SKM, Muller S. "An Overview of Novel Adjuvants Designed for Improving Vaccine Efficacy". Trends Pharmacol Sci. 2017, 38(9): pp. 771–793.
- [72]. Erdei L, Bolla BS, Bozó R, Tax G, Urbán E, Kemény L, et al. "TNIP1 regulates Cutibacterium acnes-induced innate immune functions in epidermal keratinocytes". Front Immunol. 2018, 9(9): pp. 1–11.
- [73]. Alandijany T, Roberts APE, Conn KL, Loney C, McFarlane S, Orr A, et al. "Distinct temporal roles for the promyelocytic leukaemia (PML) protein in the sequential regulation of intracellular host immunity to HSV-1 infection". PLoS Pathog. 2018, 14(2): pp. 1–36.
- [74]. McFarlane S, Orr A, Roberts APE, Conn KL, Iliev V, Loney C, et al. "The histone chaperone hira promotes the induction of host innate immune defences in response to HSV-1 infection". PLoS Pathog. 2019, 15(3): pp. 1– 36.
- [75]. Ashkar AA, Rosenthal KL. "Interleukin-15 and Natural Killer and NKT Cells Play a Critical Role in Innate Protection against Genital Herpes Simplex Virus Type 2 Infection". J Virol. 2003, 77(18): pp. 10168–71.
- [76]. Gill PG, Morris PJ, Kettlewell M. "The complications of intravenous Corynebacterium parvum infusion". Clin Exp Immunol. 1977, 30, pp. 229–232.
- [77]. Ribas G, Neville M, Wixon JL, Cheng J, Campbell RD. "Alleviation of Lipopolysaccharide-Induced Acute Liver Injury in Propionibacterium acnes-Primed IFN-γ-Deficient Mice by a concomitant reduction of TNF-α IL-12, and IL-18 Production". J Immunol. 1999, 162, pp. 1049–1055.
- [78]. B. M. E. Von Blomberg, J. Glerum, J. J. Crolest, J. Stam, and H. A. Drexhage. "Harmful effects of I.V. Corynebacterium parvum given at the same time as cyclophosphamide in patients with squamous-cell carcinoma of the bronchus." Br. J. Cancer. 1980, 41, pp. 609–617.
- [79]. Mussalem JS, Squaiella-Baptistão CC, Teixeira D, Yendo TM, Thies FG, Popi AF, et al. "Adjuvant effect of killed Propionibacterium acnes on mouse peritoneal B-1 lymphocytes and their early phagocyte differentiation". PLoS One. 2012, 7(3): pp. 1–12.
- [80]. Alexander C. Zingher D. "Propioniba cterium a cnes". J Equine Vet Sci. 1996, 16(3): pp. 100–103.
- [81]. Moorthy K. "Pharmaceutical Calculations". In: William & Wilkins, Baltimore editor. Fundamentals of Biochemical Calculations. 11th ed. 2020. pp. 173–178.