Smut Fungi: A Comprehensive Review

Sudhir Diwase¹ Department of Botany, Vasantrao Naik Mahavidyalaya, Aurangabad-431 003, India

Balasaheb Ughade³ Department of Botany, Amdar Shashikant Shinde Mahavidyalaya, Medha, India

Tejswini Sontakke⁵ Department of Zoology, MGV's, MPH Mahila College, Malegaon, Dist. Nashik (MH), India

Abstract:- Smut fungi (Ustilaginales), obligate plant pathogens within the Basidiomycota phylum, play critical roles in ecology and agriculture. Known for their host specificity and production of black powdery teliospores, these fungi predominantly infect monocots, including economically vital crops like wheat, rice, maize, and sugarcane. Their global distribution and adaptability to diverse climates make them significant contributors to agricultural losses, impacting food security and trade. Traditional methods for identifying smut fungi, such as morphological and cultural analyses, are now complemented by molecular techniques like DNA barcoding, PCR, and next-generation sequencing (NGS). These advancements have refined fungal taxonomy, uncovered cryptic species, and elucidated evolutionary relationships, enhancing the accuracy of identification and ecological understanding. India, a biodiversity hotspot, reports 159 species of smut fungi, primarily targeting the Poaceae family. Despite advancements, gaps remain in understanding their biodiversity, pathogenic mechanisms, and responses to climate change. Addressing these challenges necessitates interdisciplinary research, integrating modern molecular tools with traditional approaches. This review underscores the importance of smut fungi research for developing sustainable disease management strategies. By fostering global collaboration and leveraging advanced techniques, researchers can mitigate the agricultural impact of smut fungi while exploring their ecological and biotechnological potential. Comprehensive studies are crucial for ensuring agricultural sustainability, biodiversity conservation, and enhanced food security in the face of emerging global challenges.

Keywords:- Smut Fungi, Fungal Taxonomy, Plant Pathogens, Molecular Identification, Agricultural Sustainability. Arvind Dhabe² Department of Botany, Dr. B. A. M. University, Aurangabad-431 004, India

Ashwini Biradar⁴ Department of Microbiology, Dr. B. A. M. University, Sub Campus Osmanabad, Osmanabad 413501, India

Dinesh Nalage⁶ Department of Biotechnology, Maulana Azad College of Arts, Science & Commerce, Aurangabad 431001 (MH), India.

I. INTRODUCTION

Fungi represent one of the most significant components of Earth's biotic system, contributing immensely to ecological balance, biodiversity, and agricultural systems. An estimated 1.5 million fungal species have evolved over approximately one billion years; however, only about 5% of these species have been documented and explored, leaving 95% of fungal diversity unknown and uncharacterized (Vabeikhokhei et al., 2019). This massive gap underscores the importance of advancing research in the field of mycology, particularly in regions with high biodiversity such as the tropics and subtropics.

Fungi are eukaryotic microorganisms that thrive in diverse ecosystems across the globe, including extreme environments like the Arctic, Antarctic, African deserts, and deep-sea thermal vents (Mikryukov et al., 2023; Tedersoo et al., 2014). Their versatility stems from their ability to adapt to diverse climatic and ecological conditions, playing key roles in nutrient cycling, decomposition, and symbiotic relationships with plants and animals.

II. SYSTEMATIC CLASSIFICATION AND DIVERSITY OF FUNGI

Fungal taxonomy has been refined through morphological, biochemical, and molecular studies, leading to a clearer understanding of their diversity and evolutionary history. Fungi are traditionally classified into four major phyla:

- Chytridiomycota: Aquatic fungi with flagellated spores, often found in water and soil environments.
- Zygomycota: Terrestrial fungi known for their role in food spoilage and industrial fermentation.
- Ascomycota: The largest fungal group, comprising yeasts, molds, and filamentous fungi.
- Basidiomycota: Includes macroscopic fungi such as mushrooms, bracket fungi, rust fungi, and smut fungi.

Volume 9, Issue 12, December – 2024

ISSN No:-2456-2165

Recent advancements in molecular techniques have uncovered new fungal lineages, leading to the identification of emerging fungal groups that challenge the traditional taxonomic hierarchy.

In the Indian context, fungal diversity is unparalleled, with more than 27,000 species identified to date. India ranks among the top biodiversity hotspots, with its diverse climatic zones fostering the growth of fungi across 103 orders, 484 families, and 4,979 genera(Punjabi et al., 2020; Singh, 2020).

III. BASIDIOMYCOTA: THE ROLE OF SMUT FUNGI

Within the phylum Basidiomycota, basidiomycetes represent approximately 30% of the total fungal diversity. This group includes smut fungi, rust fungi, and other plant pathogens that hold immense ecological and economic significance. Smut fungi alone constitute about 10% of the Basidiomycota, with their characteristic black powdery spores forming on infected plant tissues.

Smut fungi are obligate plant-parasitic microorganisms within the order Ustilaginales. They are second only to rust fungi in terms of species diversity and impact. These fungi are predominantly known for infecting monocots, particularly members of the Poaceae (grasses) and Cyperaceae (sedges) families, although they also parasitize dicots and herbaceous plants.

➤ Key Features of Smut Fungi:

- Production of black powdery spores (teliospores) on infected tissues.
- Obligate biotrophic lifecycle, requiring a living host for completion.
- Infection primarily targets reproductive organs (e.g., flowers, seeds) and meristematic tissues.
- Widespread presence in both temperate and tropical climates.

IV. GLOBAL DISTRIBUTION AND IMPORTANCE OF SMUT FUNGI

Globally, smut fungi are distributed across all continents, with notable prevalence in temperate regions where conditions such as 30°C temperature and 100% relative humidity favor their proliferation(Gautam et al., 2021). The ability of smut fungi to adapt to diverse ecological conditions contributes to their persistence and impact on agricultural systems. For example:

- Ustilago cynodontis (Couch smut): Infects *Cynodon dactylon* (turfgrass) and thrives in temperate climates like Australia and desert environments such as Egypt(García-Guzmán & Burdon, 1997).
- Ustilago sporoboli-indici: Successfully invaded southeast Queensland in Australia, where it infects *Sporobolus natalensis*. Originally endemic to Africa and Asia, its rapid spread exemplifies the invasive potential of smut fungi(Steinrucken & Vitelli, 2023).

V. INDIAN CONTEXT: DIVERSITY AND DISTRIBUTION OF SMUT FUNGI

https://doi.org/10.5281/zenodo.14608940

India's unique geographic and agro-climatic diversity makes it an ideal location for studying fungal biodiversity. The country harbors eight out of twelve recognized climatic zones, ranging from temperate to alpine regions, with significant variations in temperature, rainfall, and humidity. These conditions support the growth of a diverse range of smut fungi, particularly those parasitizing economically significant crops.

From an Indian Perspective:

- 159 species of smut fungi belonging to 18 genera have been reported.
- These species parasitize 189 host plants spanning eight families, with Poaceae being the most affected(Gautam et al., 2021).
- The genus Ustilago dominates Indian smut fungi, comprising 48 species (30.38% of the total reported species). Other important genera include Sporisorium and Anthracocystis.
- Lesser-Known Genera in India:
- Ahmadiago
- Bambusiomyces
- Cintractia
- Clinoconidium
- Farysia
- Macalpinomyces
- Melanopsichium
- Moesziomyces
- Pericladium

These genera often exhibit host specificity, targeting specific crop species and ecosystems.

VI. MODE OF INFECTION AND LIFE CYCLE OF SMUT FUNGI

Smut fungi are systemic pathogens that target specific plant tissues, primarily reproductive organs. The life cycle of smut fungi involves:

> Teliospore Production:

Smut fungi produce teliospores (thick-walled, melanized spores) on infected plant tissues. These spores serve as survival structures under adverse environmental conditions.

Spore Germination:

Under favorable conditions, teliospores germinate, producing basidiospores that initiate infection.

> Infection Process:

Basidiospores infect host tissues, leading to systemic colonization. Infection often begins in the seedling stage and progresses to the reproductive organs.

Volume 9, Issue 12, December – 2024

ISSN No:-2456-2165

➤ Host Tissue Modification:

Infected tissues develop tumors or galls filled with fungal spores. For example, Ustilago maydis causes tumor formation in maize kernels.

Spore Dissemination:

Mature spores are released into the environment, completing the lifecycle and perpetuating infection.

Unique Features of Smut Fungal Infection:

- Smut fungi utilize effector proteins to suppress host immunity and manipulate plant cellular processes.
- They exhibit host specificity, infecting selected plant species while remaining dormant in others.

VII. IMPACT OF SMUT FUNGI ON AGRICULTURE

Smut fungi pose a significant threat to global agriculture by reducing crop yield, quality, and economic value. Their ability to infect staple crops like wheat, rice, maize, sugarcane, and sorghum exacerbates challenges related to food security and trade restrictions.

➤ Wheat Smuts

Wheat, a vital cereal crop grown on 215 million hectares worldwide, is highly susceptible to smut fungi such as:

- Tilletia indica: Causes Karnal bunt.
- Tilletia tritici and T. laevis: Cause loose smut and stinking bunt, respectively.

Although Karnal bunt causes yield losses of only 0.5%, its economic impact is disproportionate due to quarantine regulations in Europe and the US (Turgay et al., 2020). In Germany, estimated financial losses due to Karnal bunt exceed $\notin 5$ million, highlighting the trade-related consequences of smut diseases.

➢ Rice Smuts

Rice (*Oryza sativa*), a staple food for over half the world's population, is vulnerable to:

- Rice False Smut (*Ustilaginoidea virens*): Yield losses exceed 75% in heavily affected regions.
- Rice Karnal Smut (*Tilletia horrida*): Incidence rates reach 100% in China and 87% in Pakistan(Bishnoi et al., 2020).
- ➢ Corn Smut

Corn smut, caused by Ustilago maydis, infects maize (*Zea mays*), leading to tumor-like galls on kernels and stems. Despite its destructive potential, corn smut has gained economic significance as a culinary delicacy known as "huitlacoche" in Mexico.

➤ Sugarcane Smut

Sugarcane (*Saccharum officinarum*), a major cash crop, suffers devastating yield losses from sugarcane smut caused by *Ustilago scitaminea* (syn. *Sporisorium scitamineum*).

Under severe conditions, losses may reach up to 80%, with sugar content reduced by 20%.

https://doi.org/10.5281/zenodo.14608940

VIII. HUMAN AND ANIMAL HEALTH IMPLICATIONS

While smut fungi primarily infect plants, their impact on human and animal health cannot be overlooked. The rice false smut pathogen Ustilaginoidea virens produces toxic metabolites such as ustiloxins and ustilaginoidins, which cause:

- Hepatotoxicity: Liver damage in humans and animals.
- Teratogenesis: Disruption of embryonic development in pregnant individuals(Zhou et al., 2024).
- Identification Techniques for Smut Fungi
- A. Conventional Techniques

> Morphological Analysis

Traditionally, morphological analysis has been the cornerstone of fungal identification. This approach relies on the examination of physical characteristics such as spore shape, size, ornamentation, coloration, hyphal structures, and reproductive organs(G. Fischer & Dott, 2002; Lücking et al., 2020). For smut fungi, the teliospore is often the most diagnostic structure, and taxonomists have long depended on microscopic observations to differentiate species.

- Advantages:
- ✓ Simplicity and Cost-Effectiveness: Morphological analyses require only basic tools, such as a compound microscope and simple staining techniques. This makes them accessible and relatively inexpensive, which is important for laboratories with limited resources.
- ✓ Direct Examination of Material: The immediate observation of infected plant material and fungal structures in situ provides a direct link between the pathogen and the symptoms it causes, aiding in initial diagnoses.
- Limitations:
- ✓ Phenotypic Plasticity: Morphological traits can vary significantly depending on environmental conditions, host species, and the developmental stage of the fungus. Such phenotypic plasticity often leads to misidentification and confusion, especially when distinguishing closely related species.
- ✓ Subjectivity: Interpreting morphological characteristics can be subjective, relying heavily on the expertise and experience of the mycologist. Minor differences in spore dimensions or hyphal structures can be overlooked or misread.

Despite these limitations, morphological analysis remains a foundational technique and continues to be valuable for preliminary identification and for guiding subsequent, more definitive methods.

Volume 9, Issue 12, December – 2024

ISSN No:-2456-2165

➤ Cultural Techniques

Cultural techniques involve isolating the fungus from infected tissues and growing it on artificial media. By doing this, researchers can observe colony morphology, growth rates, pigmentation, and other cultural characteristics under controlled conditions(Afifah & Saputro, 2020; Humber, 2012; Mendoza-martínez et al., 2020; Singh & Sharma, 2020; Uikey et al., 2020).

- Advantages:
- ✓ Isolation and Maintenance of Pure Cultures: Culturing the fungus allows the development of stable reference strains, essential for further studies, including genetic analyses and pathogenicity tests.
- ✓ Characterization of Growth Traits: Colony characteristics, such as texture, growth rate, and color, can provide supplementary clues for identification.
- Limitations:
- ✓ Obligate Biotrophy: Many smut fungi are obligate biotrophs, meaning they rely on a living host for nutrition and may not grow readily on artificial media. This makes culturing challenging or impossible for certain species.
- ✓ Overlapping Morphologies: Even if a fungus grows in culture, variations in nutrient media and conditions can lead to phenotypic changes that are not reliable for accurate species-level identification.

Cultural techniques, while useful, are often employed as a complement to morphological and molecular methods, rather than serving as a standalone identification tool.

➢ Biochemical Tests

Biochemical profiling involves assessing enzyme activities, metabolic products, and other physiological traits that can help differentiate species. Some smut fungi may exhibit distinct enzyme activity patterns or utilize certain substrates more efficiently, providing a biochemical "fingerprint" for identification(Jaswal et al., 2020; Schirawski et al., 2021; Steins et al., 2023; Uikey et al., 2020; Xia et al., 2020).

- Advantages:
- ✓ Additional Data Layer: Biochemical tests add another dimension to classification, helping distinguish species that appear morphologically similar.
- ✓ Metabolic Insights: Understanding the metabolic capabilities of a fungus can shed light on its ecological role and host interactions.
- Limitations:
- ✓ Overlap of Biochemical Traits: Many smut fungi share similar biochemical characteristics, limiting the discriminatory power of such tests. Two or more closely related species may exhibit nearly identical enzyme activity profiles.

✓ Requirement for Specialized Assays: Biochemical tests may demand specialized reagents, equipment, and expertise, adding complexity and cost to the identification process.

https://doi.org/10.5281/zenodo.14608940

As with cultural methods, biochemical tests are often integrated with other techniques to strengthen the accuracy of species identification.

> Pathosystem Studies

Pathosystem studies focus on the interaction between the fungus and its host plant. Observing disease symptoms, host specificity, and infection processes provides a deeper ecological and evolutionary context(Hu et al., 2022; Mapuranga et al., 2022; Nirmalkar et al., 2020; Sharma, 2021; Vorob'eva & Toropova, 2020).

- Advantages:
- ✓ Ecological and Biological Relevance: Studying the hostpathogen dynamics reveals information about virulence factors, infection strategies, and life cycle stages, which can complement morphological and molecular data.
- ✓ Confirming Pathogenic Identity: When smut fungi are identified based on their pathogenicity and host range, it can help differentiate them from non-pathogenic, morphologically similar fungi.
- Limitations:
- ✓ Time-Consuming: Such studies are often protracted, as the fungus must be inoculated onto host plants, and the development of disease symptoms may take weeks or months.
- ✓ Controlled Conditions Required: Maintaining appropriate environmental conditions and selecting suitable host plants can be challenging, limiting the practical feasibility of using pathosystem studies as a routine identification technique.

Pathosystem studies are invaluable for understanding the biology and ecology of smut fungi but are not typically the first line of identification due to the logistical challenges involved.

B. Molecular Techniques

With the advent of molecular biology, more precise, reliable, and rapid identification methods have emerged. Molecular techniques target DNA or RNA sequences, providing stable, heritable markers that are less influenced by environmental factors and phenotypic plasticity(Haddrill, 2021; Hasnain & Mehvish, 2020; Kartavtsev, 2021; Kasper et al., 2020; Kumawat et al., 2021; Yali, 2022).

> DNA Barcoding and Sequencing

DNA barcoding involves using standardized genomic regions as universal identifiers. In fungi, the Internal Transcribed Spacer (ITS) region of the ribosomal RNA gene cluster has become the primary barcode due to its high level of interspecific variability and ease of amplification(Santamaria, 2011; Seifert, 2009).

ISSN No:-2456-2165

https://doi.org/10.5281/zenodo.14608940

- Advantages:
- ✓ High Accuracy and Reliability: ITS sequences allow for the differentiation of closely related species, including cryptic species that appear identical morphologically.
- ✓ Global Standardization: Due to the widespread use of ITS as a fungal barcode, large reference databases (e.g., GenBank, UNITE) facilitate rapid species identification.

Example Applications: Studies have employed ITS sequencing to resolve taxonomic ambiguities among smut fungi. For example, Seifert, 2009 and Fischer & Hirschhorn, 2018 demonstrated how ITS sequencing has clarified relationships within the Ustilaginaceae, revealing new lineages and correcting previous misclassifications. Such work underscores the power of DNA barcoding to refine fungal taxonomy and improve the accuracy of identification(Tiknaik et al., 2019).

> PCR-Based Methods

Polymerase Chain Reaction (PCR) techniques enable the amplification of specific DNA fragments from minute amounts of fungal tissue(Eberhardt, 2012; Hosoya et al., 2012). By designing taxon-specific primers that target unique genomic regions, PCR-based methods can rapidly confirm a fungus's identity.

- Advantages:
- ✓ Rapid and Sensitive: PCR can detect and amplify fungal DNA even from very small samples, including those obtained from infected plant tissue or environmental samples.
- ✓ Direct Application to Field Samples: Samples collected in the field can be processed without the need for culturing, accelerating the identification process.

PCR techniques are often combined with DNA sequencing for definitive species identification, making them a critical component of modern diagnostic and taxonomic workflows(Balajee et al., 2009; Erlich et al., 1991).

> Phylogenetic Analysis

Beyond just identifying species, molecular data enable researchers to reconstruct phylogenetic trees that illustrate evolutionary relationships. By comparing gene sequences (e.g., ITS, LSU, EF1- α) across multiple taxa, mycologists can discern patterns of divergence, speciation, and host adaptation(Castelle & Banfield, 2018; Fitzpatrick et al., 2006; McCarthy & Fitzpatrick, 2017; Rahayu et al., 2021).

- Advantages:
- ✓ Evolutionary Insights: Phylogenetic analyses help interpret how smut fungi have evolved, migrated, and adapted to different hosts or environmental conditions.
- ✓ Refinement of Taxonomy: When morphological traits conflict with traditional taxonomy, molecular phylogenies can guide the reclassification of species into more accurate lineages.

Such analyses have redefined fungal systematics, often leading to the discovery of cryptic species complexes and the rearrangement of genera and families, thereby providing a more robust taxonomic framework.

➢ Next-Generation Sequencing (NGS) and Genomic Approaches

The rapid advancement of sequencing technologies has ushered in a new era of fungal identification. Next-Generation Sequencing (NGS) techniques allow for wholegenome sequencing and metagenomic analyses that can characterize complex fungal communities, including those containing unculturable or rare species(Ahrendt et al., 2018; Araujo & Sampaio-Maia, 2018; James et al., 2020; Nalage et al., 2024; Patil et al., 2023b, 2023a; Sangal et al., 2014; Sontakke et al., 2023).

- Advantages:
- ✓ High-Throughput Identification: NGS can process thousands of samples simultaneously, making it ideal for large-scale biodiversity surveys and environmental monitoring.
- ✓ Comprehensive Genomic Insights: Whole-genome sequencing unveils genes involved in pathogenicity, host specificity, and other functional traits, thereby deepening our understanding of smut fungi biology.
- ✓ Metagenomics: By analyzing environmental DNA (eDNA) samples, NGS can detect smut fungi that are not easily cultured or observed, broadening the known diversity of these pathogens(Nalage et al., 2023; Sontakke et al., 2022).

NGS-based techniques are still relatively costly and require bioinformatics expertise. However, as these technologies become more accessible and affordable, they will likely become a standard tool for smut fungal identification and research.

IX. INTEGRATING MULTIPLE APPROACHES

Each identification technique has its strengths and weaknesses. A robust identification strategy often integrates multiple approaches, leveraging the simplicity and cost-effectiveness of morphological or cultural techniques with the precision and reliability of molecular methods(Nalage et al., 2024). For example, a field researcher might initially rely on morphological characteristics and disease symptoms to identify a suspected smut fungus. Subsequently, DNA barcoding and PCR-based tests can confirm the identification, while phylogenetic analyses and NGS data can provide deeper evolutionary and ecological insights.

This integrative strategy ensures that identifications are both accurate and informative, supporting effective disease management plans in agriculture and contributing to a more complete understanding of smut fungal diversity and evolution. ISSN No:-2456-2165

X. GAPS IN RESEARCH AND FUTURE PERSPECTIVES

Despite significant advancements in fungal taxonomy and molecular identification techniques, several gaps persist in smut fungi research. Addressing these challenges requires global collaboration, interdisciplinary approaches, and innovative technologies:

Unexplored Biodiversity:

- While smut fungi diversity has been well-studied in certain regions, large portions of the world, particularly tropical and subtropical zones, remain underexplored.
- Countries with high biodiversity, such as those in Africa, Southeast Asia, and South America, lack comprehensive surveys of smut fungi.
- > Taxonomic Ambiguity:
- Morphologically similar species and cryptic species complexes pose challenges to accurate identification.
- Integrating molecular phylogenetics with traditional taxonomy can resolve ambiguities and refine fungal classification.
- Host-Pathogen Interaction Mechanisms:
- Limited understanding of the molecular mechanisms governing host specificity, effector proteins, and fungal adaptation to host plants.
- Research into fungal effectors and their role in host immune suppression is critical for developing targeted control strategies.
- > Impact of Climate Change:
- Climate change is altering fungal distribution and pathogenicity, yet little research has been conducted on how these changes impact smut fungi.
- Predictive models and field studies are needed to assess future risks to global agriculture.
- ➢ Biotechnological Applications:
- Smut fungi, particularly *Ustilago maydis*, have shown potential in industrial applications such as bioconversion and bioremediation.
- Further research can unlock their utility in sustainable agriculture and industrial biotechnology.
- Integrated Disease Management:
- Current control measures rely heavily on chemical fungicides, which pose environmental and health concerns.
- Developing eco-friendly strategies such as biological control agents, resistant crop varieties, and cultural practices remains a priority.

- Global Research Collaboration:
- Addressing these gaps requires international cooperation, data sharing, and joint research initiatives.

https://doi.org/10.5281/zenodo.14608940

• Platforms such as global fungal databases and research networks can facilitate collaboration among scientists worldwide.

XI. CONCLUSION

Smut fungi, as obligate plant pathogens, hold significant ecological, economic, and scientific importance. While they pose substantial threats to global agriculture by reducing crop yields and quality, they also offer opportunities to advance scientific knowledge and biotechnological applications. Despite notable progress in fungal taxonomy and molecular identification, large gaps remain in understanding their biodiversity, pathogenic mechanisms, and environmental impacts. The need for interdisciplinary research and global collaboration cannot be overstated. By combining traditional methods with modern molecular techniques, researchers can unlock the full potential of smut fungi studies. Addressing these gaps will not only enhance disease management strategies but also contribute to broader ecological and evolutionary research.As climate change and emerging agricultural challenges continue to shape the future, comprehensive research on smut fungi will be essential for ensuring food security, protecting biodiversity, and fostering sustainable agricultural practices. The identification of smut fungi has evolved from a reliance on traditional, morphologybased methods to a more holistic approach that incorporates molecular and genomic techniques. Although conventional methods remain valuable for initial screenings and certain diagnostic contexts, DNA barcoding, PCR-based techniques, phylogenetic analyses, and next-generation sequencing have revolutionized fungal taxonomy, delivering higher resolution, greater accuracy, and deeper insights into the biology and diversity of these pathogens. As these advanced techniques become more accessible and user-friendly, the field of smut fungal identification will continue to progress, facilitating more effective disease management strategies, guiding quarantine measures, and enhancing our overall understanding of fungal biodiversity and evolution. In this dynamic landscape, the integration of multiple identification techniques is key, ensuring that taxonomic conclusions are well-grounded and that the resulting knowledge can be applied for the benefit of agricultural sustainability, biosecurity, and fundamental mycological research.

REFERENCES

[1]. Afifah, L., & Saputro, N. W. (2020). Growth and viability of entomopathogenic fungus Beauveria bassiana (Balsamo) Vuillemin in different alternative media. IOP Conference Series: Earth and Environmental Science, 468(1), 012037. https://doi.org/10.1088/1755-1315/468/1/012037

https://doi.org/10.5281/zenodo.14608940

- [2]. Ahrendt, S. R., Quandt, C. A., Ciobanu, D., Clum, A., Salamov, A., Andreopoulos, B., Cheng, J.-F., Woyke, T., Pelin, A., Henrissat, B., Reynolds, N. K., Benny, G. L., Smith, M. E., James, T. Y., & Grigoriev, I. V. (2018). Leveraging single-cell genomics to expand the fungal tree of life. Nature Microbiology, 3(12), 1417– 1428. https://doi.org/10.1038/s41564-018-0261-0
- [3]. Araujo, R., & Sampaio-Maia, B. (2018). Fungal Genomes and Genotyping. In Advances in Applied Microbiology (Vol. 102, pp. 37–81). Elsevier. https://doi.org/10.1016/bs.aambs.2017.10.003
- [4]. Balajee, S. A., Borman, A. M., Brandt, M. E., Cano, J., Cuenca-Estrella, M., Dannaoui, E., Guarro, J., Haase, G., Kibbler, C. C., Meyer, W., O'Donnell, K., Petti, C. A., Rodriguez-Tudela, J. L., Sutton, D., Velegraki, A., & Wickes, B. L. (2009). Sequence-Based Identification of Aspergillus, Fusarium, and Mucorales Species in the Clinical Mycology Laboratory: Where Are We and Where Should We Go from Here? Journal of Clinical Microbiology, 47(4), 877–884. https://doi.org/10.1128/JCM.01685-08
- [5]. Bishnoi, S. K., He, X., Phuke, R. M., Kashyap, P. L., Alakonya, A., Chhokar, V., Singh, R. P., & Singh, P. K. (2020). Karnal Bunt: A Re-Emerging Old Foe of Wheat. Frontiers in Plant Science, 11(September), 1– 18. https://doi.org/10.3389/fpls.2020.569057
- [6]. Castelle, C. J., & Banfield, J. F. (2018). Major New Microbial Groups Expand Diversity and Alter our Understanding of the Tree of Life. Cell, 172(6), 1181– 1197. https://doi.org/10.1016/j.cell.2018.02.016
- [7]. Eberhardt, U. (2012). Methods for DNA Barcoding of Fungi. In W. J. Kress & D. L. Erickson (Eds.), DNA Barcodes (Vol. 858, pp. 183–205). Humana Press. https://doi.org/10.1007/978-1-61779-591-6_9
- [8]. Erlich, H. A., Gelfand, D., & Sninsky, J. J. (1991). Recent Advances in the Polymerase Chain Reaction. Science, 252(5013), 1643–1651. https://doi.org/10.1126/science.2047872
- [9]. Fischer, G., & Dott, W. (2002). Quality assurance and good laboratory practice in the mycological laboratory – compilation of basic techniques for the identification of fungi. International Journal of Hygiene and Environmental Health, 205(6), 433–442. https://doi.org/10.1078/1438-4639-00190
- [10]. Fischer, G. W., & Hirschhorn, E. (2018). A Critical Study of Some Species of Ustilago Causing Stem Smut on Various Grasses. Mycologia, 37(2), 236–266. https://doi.org/10.1080/00275514.1945.12023984
- [11]. Fitzpatrick, D. A., Logue, M. E., Stajich, J. E., & Butler, G. (2006). A fungal phylogeny based on 42 complete genomes derived from supertree and combined gene analysis. BMC Evolutionary Biology, 6(1), 99. https://doi.org/10.1186/1471-2148-6-99
- [12]. García-Guzmán, G., & Burdon, J. J. (1997). Impact of the flower smut Ustilago cynodontis (Ustilaginaceae) on the performance of the clonal grass Cynodon dactylon (Gramineae). American Journal of Botany, 84(11), 1565–1571. https://doi.org/10.2307/2446618

- [13]. Gautam, A. K., Verma, R. K., Avasthi, S., Sushma, S., Devadatha, B., Thakur, S., Kashyap, P. L., Prasher, I. B., Bhadauria, R., Niranjan, M., & Ranadive, K. R. (2021). Smut fungi: A compendium of their diversity and distribution in India. MycoAsia. https://doi.org/10.59265/mycoasia.2021-01
- [14]. Haddrill, P. R. (2021). Developments in forensic DNA analysis. Emerging Topics in Life Sciences, 5(3), 381–393. https://doi.org/10.1042/ETLS20200304
- [15]. Hasnain, H., & Mehvish, N. (2020). Assessment of plant genetic variations using molecular markers: A review. Journal of Applied Biology & Biotechnology. https://doi.org/10.7324/JABB.2020.80514
- [16]. Hosoya, K., Nakayama, M., Matsuzawa, T., Imanishi, Y., Hitomi, J., & Yaguchi, T. (2012). Risk analysis and development of a rapid method for identifying four species of Byssochlamys. Food Control, 26(1), 169–173.

https://doi.org/10.1016/j.foodcont.2012.01.024

- [17]. Hu, Y., Irinyi, L., Hoang, M. T. V., Eenjes, T., Graetz, A., Stone, E. A., Meyer, W., Schwessinger, B., & Rathjen, J. P. (2022). Inferring Species Compositions of Complex Fungal Communities from Long- and Short-Read Sequence Data. mBio, 13(2), e02444-21. https://doi.org/10.1128/mbio.02444-21
- [18]. Humber, R. A. (2012). Preservation of entomopathogenic fungal cultures. In Manual of Techniques in Invertebrate Pathology (pp. 317–328). Elsevier. https://doi.org/10.1016/B978-0-12-386899-2.00010-5
- [19]. James, T. Y., Stajich, J. E., Hittinger, C. T., & Rokas, A. (2020). Toward a Fully Resolved Fungal Tree of Life. Annual Review of Microbiology, 74(1), 291– 313. https://doi.org/10.1146/annurev-micro-022020-051835
- [20]. Jaswal, R., Rajarammohan, S., Dubey, H., & Sharma, T. R. (2020). Smut fungi as a stratagem to characterize rust effectors: Opportunities and challenges. World Journal of Microbiology and Biotechnology, 36(10), 150. https://doi.org/10.1007/s11274-020-02927-x
- [21]. Kartavtsev, Y. P. (2021). Some Examples of the Use of Molecular Markers for Needs of Basic Biology and Modern Society. Animals, 11(5), 1473. https://doi.org/10.3390/ani11051473
- [22]. Kasper, C., Ribeiro, D., Almeida, A. M. D., Larzul, C., Liaubet, L., & Murani, E. (2020). Omics Application in Animal Science—A Special Emphasis on Stress Response and Damaging Behaviour in Pigs. Genes, 11(8), 920. https://doi.org/10.3390/genes11080920
- [23]. Kumawat, G., Kanta Kumawat, C., Chandra, K., Pandey, S., Chand, S., Nandan Mishra, U., Lenka, D., & Sharma, R. (2021). Insights into Marker Assisted Selection and Its Applications in Plant Breeding. In I. Y. Abdurakhmonov (Ed.), Plant Breeding—Current and Future Views. IntechOpen. https://doi.org/10.5772/intechopen.95004

- [24]. Lücking, R., Aime, M. C., Robbertse, B., Miller, A. N., Ariyawansa, H. A., Aoki, T., Cardinali, G., Crous, P. W., Druzhinina, I. S., Geiser, D. M., Hawksworth, D. L., Hyde, K. D., Irinyi, L., Jeewon, R., Johnston, P. R., Kirk, P. M., Malosso, E., May, T. W., Meyer, W., ... Schoch, C. L. (2020). Unambiguous identification of fungi: Where do we stand and how accurate and precise is fungal DNA barcoding? IMA Fungus, 11(1), 14. https://doi.org/10.1186/s43008-020-00033-z
- [25]. Mapuranga, J., Zhang, N., Zhang, L., Chang, J., & Yang, W. (2022). Infection Strategies and Pathogenicity of Biotrophic Plant Fungal Pathogens. Frontiers in Microbiology, 13, 799396. https://doi.org/10.3389/fmicb.2022.799396
- [26]. McCarthy, C. G. P., & Fitzpatrick, D. A. (2017). Multiple Approaches to Phylogenomic Reconstruction of the Fungal Kingdom. In Advances in Genetics (Vol. 100, pp. 211–266). Elsevier. https://doi.org/10.1016/bs.adgen.2017.09.006
- [27]. Mendoza-martínez, A. E., Cano-domínguez, N., & Aguirre, J. (2020). Yap1 homologs mediate more than the redox regulation of the antioxidant response in fi lamentous fungi. Fungal Biology, 124(5), 253–262. https://doi.org/10.1016/j.funbio.2019.04.001
- [28]. Mikryukov, V., Dulya, O., Zizka, A., Bahram, M., Hagh-Doust, N., Anslan, S., Prylutskyi, O., Delgado-Baquerizo, M., Maestre, F. T., Nilsson, H., Pärn, J., Öpik, M., Moora, M., Zobel, M., Espenberg, M., Mander, Ü., Khalid, A. N., Corrales, A., Agan, A., ... Tedersoo, L. (2023). Connecting the multiple dimensions of global soil fungal diversity. Science Advances, 9(48), eadj8016. https://doi.org/10.1126/sciadv.adj8016
- [29]. Nalage, D., Kale, R., Sontakke, T., Pradhan, V., Biradar, A., Senevirathna, J. D. M., Jaweria, R., Dighe, T., Dixit, P., Patil, R., & Kudnar, P. S. (2024). Bacterial phyla: Microbiota of kingdom animalia. Academia Biology, 2(4). https://doi.org/10.20935/AcadBiol7423
- [30]. Nalage, D., Kudnar, P. S., Sontakke, T., Chittapure, I., Gowda, Y., Kharbal, S., & Alamwar, Y. (2024). Assessment of the status of Spodoptera species (Lepidoptera: Noctuidae: Armyworm) in India through DNA barcoding technique. Journal of Threatened Taxa, 16(7), 25528–25535. https://doi.org/10.11609/jott.8983.16.7.25528-25535
- [31]. Nalage, D., Sontakke, T., Biradar, A., Jogdand, V., Kale, R., Harke, S., Kale, R., & Dixit, P. (2023). The impact of environmental toxins on the animal gut microbiome and their potential to contribute to disease. Elsevier, 3(C).
- [32]. Nirmalkar, V. K., Lakplae, N., & Tiwari, R. K. S. (2020). Natural Occurrence and Distribution of Entomopathogenic Fungi from Chhattisgarh. International Journal of Current Microbiology and Applied Sciences, 9(1), 1990–1998. https://doi.org/10.20546/ijcmas.2020.901.225

- [33]. Patil, R., Satpute, R., & Nalage, D. (2023a). Plant microbiomes and their role in plant health. Microenvironment and Microecology Research, 5(1), 2. https://doi.org/10.53388/MMR2023002
- [34]. Patil, R., Satpute, R., & Nalage, D. (2023b). The application of omics technologies to toxicology. Toxicology Advances, 5(2), 6. https://doi.org/10.53388/TA202305006
- [35]. Punjabi, G., Jayadevan, A., Jamalabad, A., Velho, N., Niphadkar-Bandekar, M., Baidya, P., Jambhekar, R., Rangnekar, P., Dharwadkar, O., Lopez, R., Rodrigues, M., Patel, F. D., Chandra Sagar, H. S. S., Banerjee, S., Chandi, M., Mehrotra, N., Srinivasan, S., Shahi, S., Atkore, V., ... Borkar, M. R. (2020). On the inadequacy of environment impact assessments for projects in Bhagwan Mahavir Wildlife Sanctuary and National Park of Goa, India: A peer review. Journal of Threatened Taxa, 12(18), 17387–17454. https://doi.org/10.11609/jott.6650.12.18.17387-17454
- [36]. Rahayu, D. A., Ambarwati, R., & Faizah, U. (2021). An effort to train the biological computation skill and teach animal phenetic taxonomy to pre-service biology teacher. Journal of Physics: Conference Series, 1747(1), 012001. https://doi.org/10.1088/1742-6596/1747/1/012001
- [37]. Sangal, V., Nieminen, L., Tucker, N. P., & Hoskisson, P. A. (2014). Revolutionizing Prokaryotic Systematics Through Next-Generation Sequencing. In Methods in Microbiology (Vol. 41, pp. 75–101). Elsevier. https://doi.org/10.1016/bs.mim.2014.07.001
- [38]. Santamaria, M. (2011). DNA barcoding of toxigenic fungi: A perspective. In Determining Mycotoxins and Mycotoxigenic Fungi in Food and Feed (pp. 349–356). Elsevier. https://doi.org/10.1533/9780857090973.4.349
- [39]. Schirawski, J., Perlin, M. H., & Saville, B. J. (2021).
 Smuts to the power of three: Biotechnology, biotrophy, and basic biology. Journal of Fungi, 7(8), 0–3. https://doi.org/10.3390/jof7080660
- [40]. Seifert, K. A. (2009). Progress towards DNA barcoding of fungi. Molecular Ecology Resources, 9(SUPPL. 1), 83–89. https://doi.org/10.1111/j.1755-0998.2009.02635.x
- [41]. Sharma, I. (2021). Phytopathogenic fungi and their biocontrol applications. In Fungi Bio-Prospects in Sustainable Agriculture, Environment and Nano-Technology (pp. 155–188). Elsevier. https://doi.org/10.1016/B978-0-12-821394-0.00007-X
- [42]. Singh, P. (2020). Floristic Diversity of India: An Overview. In G. H. Dar & A. A. Khuroo (Eds.), Biodiversity of the Himalaya: Jammu and Kashmir State (Vol. 18, pp. 41–69). Springer Singapore. https://doi.org/10.1007/978-981-32-9174-4_3
- [43]. Singh, P., & Sharma, M. (2020). Cultural and Morphological Characterization of Antagonistic Trichoderma Isolates. International Journal of Current Microbiology and Applied Sciences, 9(3), 1041–1048. https://doi.org/10.20546/ijcmas.2020.903.122

https://doi.org/10.5281/zenodo.14608940

- ISSN No:-2456-2165
- [44]. Sontakke, T., Biradar, A., Dixit, P., & Nalage, D. (2022). Metagenomics and microbiome of infant: Old and recent instincts. Microenvironment and Microecology Research, 4(2), Article 2. https://doi.org/10.53388/MMR2022007
- [45]. Sontakke, T., Biradar, A., & Nalage, D. (2023). The role of genetics in determining resistance to coccidiosis in goats a review of current research and future directions. Molecular Biology Reports, 50(7), 6171–6175. https://doi.org/10.1007/s11033-023-08520-3
- [46]. Steinrucken, T. V., & Vitelli, J. S. (2023). Biocontrol of weedy Sporobolus grasses in Australia using fungal pathogens. BioControl, 68(4), 341–361. https://doi.org/10.1007/s10526-023-10195-5
- [47]. Steins, L., Duhamel, M., Klenner-Koch, S., Begerow, D., & Kemler, M. (2023). Resources and tools for studying convergent evolution in different lineages of smut fungi. Mycological Progress, 22(11), 76. https://doi.org/10.1007/s11557-023-01918-0
- [48]. Tedersoo, L., Bahram, M., Põlme, S., Kõljalg, U., Yorou, N. S., Wijesundera, R., Ruiz, L. V., Vasco-Palacios, A. M., Thu, P. Q., Suija, A., Smith, M. E., Sharp, C., Saluveer, E., Saitta, A., Rosas, M., Riit, T., Ratkowsky, D., Pritsch, K., Põldmaa, K., ... Abarenkov, K. (2014). Global diversity and geography of soil fungi. Science, 346(6213), 1256688. https://doi.org/10.1126/science.1256688
- [49]. Tiknaik, A., Kalyankar, A., Shingare, M., Suryawanshi, R., Prakash, B., Sontakke, T. A., Nalage, D., Sanil, R., & Khedkar, G. (2019). Refutation of media reports on introduction of the red bellied piranha and potential impacts on aquatic biodiversity in India. Mitochondrial DNA Part A, 30(4), 643–650. https://doi.org/10.1080/24701394.2019.1611798
- [50]. Turgay, E. B., Oğuz, A. Ç., & Ölmez, F. (2020). Karnal bunt (Tilletia indica) in wheat. Climate Change and Food Security with Emphasis on Wheat, 1847, 229–241. https://doi.org/10.1016/B978-0-12-819527-7.00015-7
- [51]. Uikey, K. W., Raghuwanshi, K. S., & Uikey, D. W. (2020). Influence of Culture Media on Growth, Colony Character and Sporulation of Chaetomium globosum Fungus. International Journal of Current Microbiology and Applied Sciences, 9(5), 2567–2572. https://doi.org/10.20546/ijcmas.2020.905.293
- [52]. Vabeikhokhei, J. M. C., Mangaiha, Z., Zothanzama, J., & Lalrinawmi, H. (2019). Diversity Study of Wood Rotting Fungi from Two different Forests in Mizoram, India. International Journal of Current Microbiology and Applied Sciences, 8(04), 2775–2785. https://doi.org/10.20546/ijcmas.2019.804.323
- [53]. Vorob'eva, I., & Toropova, E. (2020). Fungi ecological niches of the genus Fusarium Link. BIO Web of Conferences, 24, 00095. https://doi.org/10.1051/bioconf/20202400095

- [54]. Xia, W., Yu, X., & Ye, Z. (2020). Smut fungal strategies for the successful infection. Microbial Pathogenesis, 142, 104039. https://doi.org/10.1016/j.micpath.2020.104039
- [55]. Yali, W. (2022). Molecular Markers: Their Importance, Types, and Applications in Modern Agriculture. Agriculture, Forestry and Fisheries, 11(1), 8. https://doi.org/10.11648/j.aff.20221101.12
- [56]. Zhou, L., Mubeen, M., Iftikhar, Y., Zheng, H., Zhang, Z., Wen, J., Khan, R. A. A., Sajid, A., Solanki, M. K., Sohail, M. A., Kumar, A., Massoud, E. E. S., & Chen, L. (2024). Rice false smut pathogen: Implications for mycotoxin contamination, current status, and future perspectives. Frontiers in Microbiology, 15, 1344831. https://doi.org/10.3389/fmicb.2024.1344831