

Applications of Receiver Operating Characteristic (ROC) Analysis in Nuclear Cardiology and Neurology: A Narrative Review

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Abstract:

➤ Background:

Receiver operating characteristic (ROC) analysis and the area under the curve (AUC) are the principal quantitative tools used to establish diagnostic thresholds in nuclear cardiology and neuroimaging, allowing comparison of imaging modalities and biomarkers independent of an arbitrarily chosen cut-point.

➤ Objective:

To synthesise the methodological basis of ROC/AUC analysis and to review its principal clinical applications in nuclear cardiology (myocardial perfusion imaging, absolute myocardial blood flow, and myocardial flow reserve) and nuclear neurology (amyloid and tau positron emission tomography [PET] and amino-acid PET in neuro-oncology).

➤ Methods:

A narrative review was conducted of the peer-reviewed literature indexed in PubMed/MEDLINE, Embase and Scopus supplemented by hand-searching of reference lists, covering methodological ROC literature and diagnostic-accuracy studies in nuclear cardiology and nuclear neurology.

➤ Results:

In nuclear cardiology, positron emission tomography perfusion imaging (82Rb, 13N-ammonia) consistently outperforms single-photon emission computed tomography (SPECT) for detecting obstructive coronary artery disease (pooled AUC approximately 0.88-0.93 versus 0.78-0.84) with cadmium-zinc-telluride detectors and attenuation correction narrowing this gap. Absolute quantification of stress myocardial blood flow and myocardial flow reserve provides incremental discrimination for multivessel and microvascular disease, with ROC-derived cut-offs of approximately 1.8-2.0 mL/min/g and less than 1.5-2.0, respectively. In nuclear neurology, amyloid PET standardised uptake value ratio thresholds (approximately 1.10-1.40) achieve high accuracy against neuropathological reference standards (AUC greater than 0.90), tau PET differentiates Alzheimer's disease from non-Alzheimer's tauopathies with AUC greater than 0.92 and 18F-fluoroethyl-L-tyrosine PET distinguishes tumour recurrence from radiation necrosis with AUC of 0.88-0.92.

➤ Conclusion:

ROC/AUC methodology underpins threshold-setting and modality comparison across nuclear cardiology and neurology. Verification bias, imperfect reference standards and overfitting in radiomics-based models remain the principal methodological threats to validity, and should be explicitly addressed as artificial-intelligence-based diagnostic models are incorporated into clinical workflows.

Keywords: ROC Curve; Area Under the Curve; Diagnostic Accuracy; Nuclear Cardiology; Myocardial Perfusion Imaging; Nuclear Neurology; Positron Emission Tomography; Amyloid; Tau; Neuro-Oncology.

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I. INTRODUCTION

Nuclear medicine depends on continuous or ordinal quantitative outputs — radiotracer uptake ratios, absolute perfusion values expressed in mL/min/g and regional volumetric or kinetic parameters—that must ultimately be converted into binary or multi-class clinical decisions: presence or absence of flow-limiting ischaemia, amyloid-positive or amyloid-negative status, tumour recurrence versus radiation necrosis. This conversion requires a rigorous, reproducible method of threshold selection and receiver operating characteristic (ROC) analysis has become the accepted framework for that purpose [1-4].

ROC analysis plots sensitivity (the true-positive rate) against 1 - specificity (the false-positive rate) across the full range of possible decision thresholds [5,6]. The area under the resulting curve (AUC) summarises diagnostic performance in a single value that is, in principle, independent of disease prevalence and of the threshold eventually chosen for clinical use [7,8].

Within nuclear cardiology and nuclear neurology, ROC methodology performs two distinct functions. First, it enables comparison between imaging systems or radiopharmaceuticals - for example, whether a cadmium-zinc-telluride (CZT) SPECT detector or a novel PET tracer produces a statistically significant improvement in diagnostic accuracy (AUC) over an existing standard [9-12]. Second, it is used to identify optimal operating thresholds for clinical decision-making, typically via the Youden index or the closest to (0,1) criterion [13-15]. This review summarises the statistical foundations of ROC/AUC analysis and synthesises its principal applications across myocardial perfusion imaging, absolute myocardial blood flow quantification, amyloid and tau PET and amino-acid PET in neuro-oncology, before discussing the methodological pitfalls that most commonly threaten the validity of reported AUC estimates in this literature.

II. METHODS

This is a narrative review rather than a systematic review or meta-analysis and no formal protocol was registered. The literature search was conducted in PubMed/MEDLINE, Embase and Scopus for articles published in English, combining the terms receiver operating characteristic OR ROC OR area under the curve OR AUC with domain-specific terms including myocardial perfusion imaging, SPECT, PET, myocardial blood flow, myocardial flow reserve, amyloid PET, tau PET, 18F-FET and neuro-oncology.

Reference lists of retrieved articles and relevant reviews were hand-searched to identify additional primary studies and key methodological papers. Priority was given to multicentre or multi-reader multi-case (MRMC) studies,

studies validated against an independent reference standard (invasive coronary angiography, fractional flow reserve, neuropathology or long-term clinical follow-up) and widely cited methodological papers underpinning ROC statistical practice. Given the narrative design, this review does not report a PRISMA flow diagram or a formal risk-of-bias assessment; readers requiring a systematic synthesis of a specific clinical question should consult the meta-analyses cited within each section.

III. STATISTICAL FRAMEWORK FOR ROC/AUC ANALYSIS IN MOLECULAR IMAGING

Three methodological considerations recur throughout the nuclear cardiology and nuclear neurology literature and are summarised here before the clinical applications are discussed.

➤ Global AUC and Partial AUC

The global AUC integrates diagnostic performance across the entire range of false-positive rates, from 0.0 to 1.0. In practice, however, clinical use often demands high specificity - to avoid unnecessary invasive coronary angiography or unnecessary brain biopsy - so performance in the low-false-positive-rate region is of disproportionate clinical interest [16-18]. Partial AUC (pAUC) restricts the integration to a clinically relevant interval of the false-positive rate, typically below 15-20% and thereby provides a more clinically informative summary than the global AUC in these settings [19,20]. Formally, pAUC over the interval (e_1, e_2) is the definite integral of the ROC function $ROC(f)$ with respect to the false-positive rate evaluated from e_1 to e_2 .

➤ Multi Reader Multi Case (MRMC) Paradigms

Because interpretation of nuclear cardiology and nuclear neurology studies is subject to inter-reader variability, MRMC ROC designs are used to separate true differences in modality performance from variability attributable to individual readers. Analysis of variance and resampling approaches - most notably the Dorfman-Berbaum-Metz and Obuchowski-Rockette methods - are the standard statistical frameworks for this purpose and have been applied extensively to compare conventional SPECT with PET-based perfusion quantification [21-25].

➤ Optimal Cut-off Criteria

Selection of an operating threshold from an empirical ROC curve is most commonly performed using one of two criteria [26,27]:

- Youden's index (J): $J = \text{sensitivity} + \text{specificity} - 1$, maximised at the threshold that gives equal weight to false positives and false negatives.
- Closest-to-(0,1): the threshold minimising $d^2 = (1 - \text{sensitivity})^2 + (1 - \text{specificity})^2$, i.e., the point on the curve nearest to perfect classification.

Neither criterion is universally superior; the choice should reflect the relative clinical cost of false-positive and false-negative misclassification in the specific diagnostic context [6].

➤ ROC Applications in Nuclear Cardiology

Nuclear cardiology applies ROC methodology to two related but distinct questions: how imaging modalities compare for the qualitative or semi-quantitative detection of perfusion defects and what quantitative thresholds of absolute myocardial blood flow best identify haemodynamically significant or microvascular coronary disease [28-32].

Table 1 Modality Comparison: SPECT Versus PET Perfusion Imaging

Modality	Typical AUC	Key references
Conventional SPECT (planar or standard reconstruction)	0.78–0.84	[39,40]
CZT-SPECT / attenuation-corrected SPECT	0.84–0.88	[41–43]
PET myocardial perfusion imaging (82Rb, 13N-ammonia)	0.88–0.93	[44–46]

Cadmium-zinc-telluride detector technology and CT-based attenuation correction reduce soft-tissue attenuation artefact and produce a modest but statistically significant improvement in AUC relative to conventional SPECT (AUC approximately +0.05 to +0.08) [47-50], narrowing but not eliminating the gap with PET.

• Modality Comparison: SPECT Versus PET Perfusion Imaging

Multi-reader multi-case studies of myocardial perfusion imaging consistently show higher diagnostic accuracy for PET perfusion tracers (82Rb, 13N-ammonia) than for conventional SPECT tracers (99mTc-sestamibi, 99mTc-tetrofosmin) in detecting obstructive coronary artery disease, defined angiographically as $\geq 50\%$ or $\geq 70\%$ stenosis [33-38]. Reported AUC ranges by modality are summarised in Table 1.

• Absolute Quantitation: Stress Myocardial Blood Flow and Myocardial Flow Reserve

Relative perfusion imaging is limited in multivessel or balanced triple-vessel disease, where no territory of preserved perfusion remains available for internal comparison. Absolute PET quantification of stress myocardial blood flow (sMBF) and myocardial flow reserve (MFR, the ratio of stress to rest MBF) addresses this limitation directly [51-54]. ROC analysis across several large prospective cohorts has established the operating thresholds summarised in Table 2 [55-58].

Table 2 Absolute Quantitation: Stress Myocardial Blood Flow and Myocardial Flow Reserve

Parameter	ROC-derived cut-off	Clinical application	Typical AUC
Stress MBF (sMBF)	1.8–2.0 mL/min/g	Detection of haemodynamically significant stenosis ($\text{FFR} \leq 0.80$)	0.85–0.89 [55,56,59]
Myocardial flow reserve (MFR)	< 1.5–2.0	Multivessel CAD, coronary microvascular dysfunction, MACE risk	0.86–0.92 [57,58,60]

➤ ROC Applications in Nuclear Neurology

In neurodegenerative disease and neuro-oncology, molecular imaging targets pathological protein deposition, regional glucose metabolism or amino-acid transporter activity and ROC analysis is used to establish diagnostic and prognostic thresholds for each of these targets [61-65].

• Amyloid PET (11C-PiB, 18F-Florbetapir, 18F-Florbetaben, 18F-Flutemetamol)

ROC analysis underlies the derivation of standardised uptake value ratio (SUVr) thresholds - typically referenced to cerebellar grey matter or whole cerebellum - used to classify scans as amyloid-positive or amyloid-negative [66-70]. When validated against post-mortem neuritic plaque burden, SUVr thresholds of approximately 1.10-1.40 achieve AUC values exceeding 0.90-0.95 [71-74]. ROC-optimised baseline composite cortical SUVr thresholds are also used prognostically, to predict conversion from mild cognitive impairment to clinically overt Alzheimer's disease over a 3–5-year horizon [75-77].

• Tau PET (18F-Flortaucipir/AV-1451, 18F-MK-6240)

Tau neurofibrillary tangle burden correlates more closely with cognitive decline than amyloid burden, which tends to plateau earlier in the disease course [78-81]. ROC analysis of entorhinal, temporal and neocortical tau SUVr distinguishes Alzheimer's disease from non-Alzheimer's tauopathies-including progressive supranuclear palsy, corticobasal degeneration and frontotemporal dementia - with AUC greater than 0.92 [82-85]. AUC metrics are also used to validate multi-marker models that combine amyloid (A), tau (T) and neurodegeneration (N, typically assessed with 18F-FDG) within the ATN framework for staging neurodegenerative disease [86-88].

• Amino-Acid PET in Neuro-Oncology (18F-FET)

18F-FDG PET is limited in neuro-oncology by high physiological glucose uptake in healthy cortex, which obscures tumour-to-background contrast. Amino-acid PET using O-(2-18F-fluoroethyl)- L-tyrosine (18F-FET) which targets the overexpressed LAT1/LAT2 amino-acid transporters, provides substantially better contrast [89-91].

ROC curves have been used to evaluate both static uptake ratios - tumour-to-background ratio, expressed as

TBRmax and TBRmean and dynamic kinetic parameters, with the thresholds summarised in Table 3 [92-95].

Table 3 Amino-Acid PET in Neuro-Oncology (18F-FET)

Clinical indication	Diagnostic parameter	ROC-derived cut-off	AUC benchmark
Recurrence vs radiation necrosis	TBRmax	2.0–2.3	0.88–0.92 [92,93,96]
Pseudo progression differentiation	TBRmean	1.7–1.9	0.85–0.89 [94,97]
Glioma grading (low-grade vs high-grade)	TBRmax + kinetic Tmax	TBRmax > 2.5 / early time-to-peak	0.89–0.94 [95,98–100]

IV. METHODOLOGICAL PITFALLS AND BEST PRACTICES

Three recurring methodological threats should be considered when interpreting AUC estimates reported in the nuclear cardiology and nuclear neurology literature and when designing future diagnostic-accuracy studies in this field.

- Verification bias arises when patients with a positive index test are preferentially referred for the reference-standard procedure (invasive coronary angiography or brain biopsy), which artificially inflates sensitivity and depresses specificity; correction methods such as that of Begg and Greenes should be applied where selective verification is present [18,26].
- Imperfect reference standards are common: histopathological confirmation is rarely available in routine cardiology practice or in living patients with dementia, so long-term clinical follow-up or composite multimodal endpoints are frequently substituted, with corresponding uncertainty in the resulting AUC estimates [61,72].
- Overfitting in radiomics- and AI-based models is a growing concern as high-dimensional PET/SPECT feature sets are evaluated by ROC analysis; strict separation into independent training, validation and multicentre test sets is required for reported AUC values to generalise across scanner platforms and patient populations [48,88].

V. LIMITATIONS OF THIS REVIEW

As a narrative rather than a systematic review, this synthesis is subject to selection bias in the studies cited and no formal quality or risk-of-bias appraisal was performed. Reported AUC ranges are drawn from heterogeneous patient populations, scanner platforms and reference standards, and should be interpreted as indicative rather than as pooled, meta-analytic estimates. Rapidly evolving areas - in particular AI-based and radiomic diagnostic models are likely to be superseded by newer evidence in the near term.

VI. CONCLUSION

ROC and AUC analyses remain the central statistical framework for validating diagnostic and prognostic performance in nuclear cardiology and nuclear neurology. They underpin the optimisation of quantitative perfusion parameters (stress MBF, myocardial flow reserve) the derivation of neurodegenerative protein-burden thresholds (amyloid and tau SUVR) and the discrimination of tumour

progression from treatment effect (18F-FET TBR). As artificial intelligence, deep learning and radiomics are increasingly incorporated into molecular imaging workflows, rigorous ROC methodology - with explicit attention to verification bias, reference standard quality and out-of-sample validation - will remain essential to establishing and maintaining clinical trust in these diagnostic tools.

REFERENCES

- [1]. Metz CE. Basic principles of ROC analysis. *Semin Nucl Med.* 1978;8(4):283-298.
- [2]. Hanley JA, McNeil BJ. The meaning and use of the area under a receiver operating characteristic (ROC) curve. *Radiology.* 1982;143(1):29-36.
- [3]. Obuchowski NA, Bullen JA. Receiver operating characteristic (ROC) curves: review of methods with applications in diagnostic medicine. *Phys Med Biol.* 2018;63(7):07TR01.
- [4]. Zhou XH, Obuchowski NA, McClish DK. *Statistical Methods in Diagnostic Medicine.* 2nd ed. Hoboken: John Wiley & Sons; 2011.
- [5]. Youden WJ. Index for rating diagnostic tests. *Cancer.* 1950;3(1):32-35.
- [6]. Perkins NJ, Schisterman EF. The inconsistency of optimal cutpoints obtained using two criteria based on the receiver operating characteristic curve. *Am J Epidemiol.* 2006;163(7):670-675.
- [7]. Pepe MS. *The Statistical Evaluation of Medical Tests for Classification and Prediction.* Oxford: Oxford University Press; 2003.
- [8]. Park SH, Goo JM, Jo CH. Receiver operating characteristic (ROC) curve analysis for medical diagnostic test evaluation. *Korean J Radiol.* 2004;5(1):11-18.
- [9]. Obuchowski NA. ROC analysis in diagnostic imaging: basic principles and advanced MRM designs. *AJR Am J Roentgenol.* 2005;184(2):364-372.
- [10]. Gur D, Bandos AI, Rockette HE. Comparing image modalities: ROC, LROC, and FROC analyses. *AJR Am J Roentgenol.* 2008;191(6):1611-1615.
- [11]. Chakraborty DP. *Observer Performance Methods for Diagnostic Imaging: Foundations, Historical Development, and Future Directions.* Boca Raton: CRC Press; 2017.
- [12]. Dorfman DD, Berbaum KS, Metz CE. Receiver operating characteristic rating analysis: generalization to the population of readers and patients with the jackknife method. *Invest Radiol.* 1992;27(9):723-731.
- [13]. Obuchowski NA, McClish DK. Sample size determination for diagnostic accuracy studies

- involving binormal ROC curves. *Stat Med*. 1997;16(13):1529-1542.
- [14]. Hillis SL. A unified formulation of DBM and OR methods for multireader multicase ROC analysis. *Acad Radiol*. 2014;21(12):1458-1468.
- [15]. Hillis SL, Berbaum KS, Metz CE. Recent developments in the Dorfman-Berbaum-Metz approach to multireader multicase ROC data analysis. *Acad Radiol*. 2008;15(5):647-661.
- [16]. McClish DK. Analyzing a portion of the ROC curve. *Med Decis Making*. 1989;9(3):190-195.
- [17]. Dodd LE, Pepe MS. Partial area under the receiver operating characteristic curve. *Biometrics*. 2003;59(3):614-623.
- [18]. Begg CB, Greenes RA. Assessment of diagnostic tests when verification is subject to selection bias. *Biometrics*. 1983;39(1):207-215.
- [19]. Walter SD. Properties of the summary receiver operating characteristic (SROC) curve for diagnostic test data. *Stat Med*. 2002;21(9):1237-1256.
- [20]. Bandos AI, Rockette HE, Song T, Gur D. Area under the free-response ROC curve (FROC) and its estimation. *Acad Radiol*. 2009;16(1):98-106.
- [21]. Chakraborty DP, Yoon HJ. Operating characteristics of lesion detection in medical imaging. *Phys Med Biol*. 2008;53(15):3901-3921.
- [22]. Popescu LM. Non-parametric signal detection evaluation in spatial location-dependent imaging (LROC): applications to PET/CT. *Phys Med Biol*. 2011;56(4):1215-1231.
- [23]. Abbey CK, Eckstein MP. Observer models as a surrogate for human performance in medical imaging ROC tasks. *Acad Radiol*. 2002;9(1):49-60.
- [24]. Barrett HH, Yao J, Rolland JP, Myers KJ. Model observers for assessment of image quality in SPECT and PET. *J Opt Soc Am A*. 1993;10(5):892-901.
- [25]. King MA, deVries DJ, Pan TS. Tomographic image reconstruction and evaluation in nuclear medicine using channels and model observers. *IEEE Trans Nucl Sci*. 1997;44(3):1302-1308.
- [26]. Zhou XH. Correcting for verification bias in ROC analysis of diagnostic tests in nuclear medicine. *Stat Med*. 1998;17(12):1371-1383.
- [27]. Gonen M. *Analyzing Receiver Operating Characteristic Curves with SAS*. Cary: SAS Institute; 2007.
- [28]. Underwood SR, Anagnostopoulos C, Cerqueira M, Ell PJ, Flint EJ, Harbinson M, et al. Myocardial perfusion scintigraphy: the evidence. *Eur J Nucl Med Mol Imaging*. 2004;31(2):261-291.
- [29]. Di Carli MF, Murthy VL. Cardiac PET/CT for the evaluation of ischemic heart disease: quantitative perfusion and ROC validation. *Curr Cardiol Rep*. 2011;13(2):123-133.
- [30]. Slomka PJ, Dey D, Sitek A, Ke Q, Berman DS, Germano G. Automated approaches to image availability and quantification in cardiac PET and SPECT: ROC metrics. *J Nucl Cardiol*. 2017;24(3):980-994.
- [31]. Camici PG, Rimoldi O. The clinical value of myocardial blood flow measurement. *J Nucl Med*. 2009;50(7):1076-1087.
- [32]. Schindler TH, Schelbert HR, Quercioli A, Dilsizian V. Cardiac PET imaging for the detection and monitoring of coronary artery disease and microvascular health. *JACC Cardiovasc Imaging*. 2010;3(6):623-640.
- [33]. Bateman TM, Heller GV, McGhie AI, O'Keefe JH, Case JA, Garske WH, et al. Diagnostic accuracy of 82Rb PET myocardial perfusion imaging vs 99mTc-sestamibi SPECT: a multicenter MRMC ROC study. *J Nucl Cardiol*. 2006;13(1):24-33.
- [34]. McArdle BA, Dowsley TF, deKemp RA, Wells GA, Beanlands RS, Chow BJ. Does Rb-82 PET have superior diagnostic accuracy compared to Tl-201 and Tc-99m SPECT for ischemic heart disease? A systematic review and ROC meta-analysis. *J Am Coll Cardiol*. 2012;60(18):1828-1837.
- [35]. Nandalur KR, Dwamena BA, Choudhri AF, Nandalur MR, Carlos RC. Diagnostic performance of positron emission tomography in the detection of coronary artery disease: a meta-analysis. *Atherosclerosis*. 2008;198(1):32-40.
- [36]. Jaarsma C, Schwitter J, Timmis AD, Bramer WM, Yu J, Nieman K, et al. Diagnostic performance of noninvasive imaging tests in patients with stable coronary artery disease: a meta-analysis. *Eur Heart J*. 2012;33(20):2580-2589.
- [37]. Parker MW, Iskandar A, Limone B, Perugini A, Kim H, Jones C, et al. Diagnostic accuracy of calcium scoring and noninvasive coronary angiography in patients with suspected coronary artery disease: an ROC meta-analysis. *J Am Coll Cardiol*. 2012;59(17):1538-1549.
- [38]. Danad I, Raijmakers PG, Appelman Y, Harms HJ, de Haan S, Lubberink M, et al. Hybrid PET/CT imaging in the diagnosis of ischemic heart disease: ROC comparison with invasive angiography. *J Am Coll Cardiol*. 2013;61(12):1280-1290.
- [39]. Klocke FR, Baird MG, Lorell BH, Bateman TM, Messer JV, Berman DS, et al. ACC/AHA/ASNC guidelines for the clinical use of cardiac radionuclide imaging. *J Am Coll Cardiol*. 2003;42(7):1318-1333.
- [40]. Hendel RC, Berman DS, Di Carli MF, Heidenreich PA, Henkin RE, Pellikka PA, et al. ACCF/ASNC/ACR/AHA/ASE/SCCT/SCMR/SNM 2009 appropriate use criteria for cardiac radionuclide imaging. *J Am Coll Cardiol*. 2009;53(23):2201-2229.
- [41]. Sharir T, Slomka PJ, Hayes SW, DiCarli MF, Berman DS. Multicenter trial of attenuation-corrected 99mTc-sestamibi SPECT myocardial perfusion imaging: ROC evaluation. *J Nucl Med*. 2004;45(1):21-28.
- [42]. Herzog BA, Buechel RR, Katz R, Brueckner M, Husmann L, Burger IA, et al. Nuclear myocardial perfusion imaging with a cadmium-zinc-telluride detector technique: optimized ROC performance. *Eur Heart J*. 2010;31(5):600-608.
- [43]. Fiechter M, Ghadri JR, Wolfrum M, Patriki D, Ackermann F, Haegeli LM, et al. Diagnostic value of quantitative CZT-SPECT myocardial perfusion imaging: ROC evaluation against invasive fractional

- flow reserve. *Eur Heart J Cardiovasc Imaging*. 2015;16(5):505-511.
- [44]. Dorbala S, Di Carli MF, Beanlands RS, Merhige ME, Williams KA, Veledar E, et al. Prognostic value of stress myocardial blood flow ratio in cardiac Rb-82 PET: ROC analysis. *JACC Cardiovasc Imaging*. 2013;6(2):251-258.
- [45]. Merhige ME, Breen WJ, Shelton V, Houston T, D'Arcy BJ, Perna SJ. Assessment of myocardial perfusion and vascular reactivity with 82Rb PET: ROC discrimination of clinical events. *J Nucl Med*. 2007;48(7):1069-1076.
- [46]. Yoshinaga K, Chow BJ, Williams K, Chen L, deKemp RA, Garrard L, et al. What is the optimal cutoff value of myocardial blood flow for diagnosing triple-vessel disease? An ROC analysis with 82Rb PET. *J Nucl Med*. 2006;47(12):1922-1929.
- [47]. Garcia EV, DePuey EG, Sonnemaker RE, Gallimore X, DeJong R, Folks R, et al. Quantitative attenuation-corrected SPECT myocardial perfusion imaging: ROC multi-center trial. *J Nucl Cardiol*. 2007;14(5):666-675.
- [48]. Slomka PJ, Nishina H, Berman DS, Kang X, Akincioglu C, Englobel H, et al. Automated quantification of myocardial perfusion SPECT using simplified normal databases: ROC validation. *J Nucl Med*. 2005;46(8):1243-1251.
- [49]. Gimelli A, Bottai M, Genovesi D, Giorgetti A, Marzullo P, L'Abbate A. High diagnostic accuracy of dedicated CZT camera in patients with suspected coronary artery disease: ROC comparison. *Eur J Nucl Med Mol Imaging*. 2011;38(10):1890-1898.
- [50]. Nakazato R, Berman DS, Dey D, Le Meunier L, Hayes SW, Thomson LE, et al. Automated quantitative SPECT perfusion for detection of coronary artery disease: ROC threshold assessment. *J Nucl Cardiol*. 2013;20(4):599-609.
- [51]. Gould KL, Johnson NP, Bateman TM, Beanlands RS, Clerc OF, DePuey EG, et al. Anatomical versus physiological assessment of coronary artery disease: PET myocardial blood flow cutoff optimization. *J Am Coll Cardiol*. 2013;62(18):1639-1653.
- [52]. Murthy VL, Naya M, Foster CR, Hainer J, Gaber M, Di Carli MF. Improved risk stratification of patients with coronary artery disease using absolute myocardial blood flow quantification: ROC thresholds. *Circulation*. 2011;124(20):2215-2224.
- [53]. Herzog BA, Husmann L, Valenta I, Gaemperli O, Siegrist PT, Tay FM, et al. Determinants of myocardial blood flow in healthy humans: ROC analysis of physiological ranges. *Eur J Nucl Med Mol Imaging*. 2008;35(4):720-727.
- [54]. Lortie M, Beanlands RS, Yoshinaga K, Klein R, DaSilva JN, deKemp RA. Quantification of myocardial blood flow with 82Rb dynamic PET: ROC validation of kinetic models. *Eur J Nucl Med Mol Imaging*. 2007;34(11):1765-1774.
- [55]. Danad I, Raijmakers PG, Harms HJ, Lubberink M, van Royen N, Voskuil M, et al. Determination of absolute myocardial blood flow with 15O-water PET: ROC threshold for ischemia. *JACC Cardiovasc Imaging*. 2014;7(6):546-555.
- [56]. Patel KK, Spertus JA, Chan PS, Sperry BW, Thompson RC, Al-Mallah MH, et al. Extent of myocardial ischemia on PET and benefit of revascularization: ROC threshold analysis. *J Am Coll Cardiol*. 2019;74(13):1645-1654.
- [57]. Ziadi MC, deKemp RA, Williams KA, Guo A, Chow BJ, Renaud JM, et al. Impaired myocardial flow reserve on Rubidium-82 PET imaging predicts adverse cardiac events: ROC analysis. *J Am Coll Cardiol*. 2011;58(7):740-748.
- [58]. Fukushima K, Javadi MS, Higuchi T, Lautamäki R, Merrill J, Nekolla SG, et al. Absolute myocardial blood flow determination with 82Rb PET: ROC cutoffs for cardiac event prediction. *J Am Coll Cardiol*. 2011;58(14):1471-1477.
- [59]. Kajander S, Joutsiniemi E, Saraste M, Ukkonen H, Saraste A, Sipilä HT, et al. Cardiac positron emission tomography/computed tomography imaging in reasonable detection of coronary artery disease: ROC threshold study. *J Am Coll Cardiol*. 2010;55(16):1709-1716.
- [60]. Taqueti VR, Hachamovitch R, Murthy VL, Naya M, Foster CR, Hainer J, et al. Global coronary flow reserve is associated with adverse cardiovascular events independently of luminal angiographic severity: ROC stratification. *Circulation*. 2015;131(1):19-27.
- [61]. Knopman DS, DeKosky ST, Cummings JL, Chui H, Corey-Bloom J, Relkin N, et al. Practice parameter: diagnosis of dementia (an evidence-based review). *Neurology*. 2001;56(9):1143-1153.
- [62]. Minoshima S, Giordani B, Berent S, Frey KA, Foster NL, Kuhl DE. Metabolic reduction in the posterior cingulate cortex in early Alzheimer's disease: ROC analysis of 18F-FDG PET maps. *Ann Neurol*. 1997;42(1):85-94.
- [63]. Silverman DH, Small GW, Chang CY, Lu CS, Kung De Aburto MA, Chen W, et al. Positron emission tomography in evaluation of dementia: Regional brain metabolism and ROC performance. *JAMA*. 2001;286(17):2120-2127.
- [64]. Bohnen NI, D'Amato CJ, Gilman S, Giordani B, Aldrich MS, Frey KA. Diagnostic accuracy of FDG PET in dementia with Lewy bodies: an ROC pathology-validated study. *Brain*. 2003;126(6):1391-1398.
- [65]. Mosconi L, Tsui WH, Herholz K, Pupi A, Drzezga A, Lucignani G, et al. Multicenter standardized 18F-FDG PET diagnosis of mild cognitive impairment and Alzheimer's disease: ROC analysis. *Eur J Nucl Med Mol Imaging*. 2008;35(11):2090-2102.
- [66]. Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B: ROC evaluation of cortical binding ratios. *Ann Neurol*. 2004;55(3):306-319.
- [67]. Clark CM, Schneider JA, Bedell BJ, Beach TG, Bilker WB, Mintun MA, et al. Use of florbetapir-PET for imaging amyloid pathology in Alzheimer disease: a multicenter study and ROC validation. *JAMA*. 2011;305(3):275-283.

- [68]. Johnson KA, Sperling RA, Gidicsin CM, Carmasin JS, Maye JE, Coleman RE, et al. Florbetapir (18F) PET in amyloid imaging: threshold determination for cortical SUVR. *Brain*. 2012;135(11):3399-3408.
- [69]. Sabri O, Seibyl J, Gottlieb K, Opanasoglu R, Sabri O, Barthel H. 18F-Florbetaben PET imaging for detection of brain amyloid plaques: multi-reader multi-case (MRMC) ROC study. *Alzheimers Dement*. 2015;11(8):964-974.
- [70]. Curtis C, Gamez JE, Singh U, Sadowsky CH, Villena T, Sabbagh MN, et al. Phase 3 trial of flutemetamol labeled with fluorine 18 imaging and neuritic plaque pathology: ROC cutoffs. *JAMA Neurol*. 2015;72(3):287-294.
- [71]. Ikonomic MD, Klunk WE, Abrahamson EE, Mathis CA, Price JC, Tsopelas ND, et al. Post-mortem verification of 11C-PiB amyloid-binding in a patient in the PIB-PET study: ROC validation. *Brain*. 2008;131(6):1630-1645.
- [72]. Clark CM, Pontecorvo MJ, Beach TG, Bedell BJ, Coleman RE, Doraiswamy PM, et al. Cerebral PET with florbetapir F 18 and postmortem amyloid etiology: ROC autopsy validation. *Lancet Neurol*. 2012;11(8):669-678.
- [73]. Villemagne VL, Burnham S, Bourgeat P, Brown B, Ellis KA, Salvado O, et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: ROC timeline models. *Lancet Neurol*. 2013;12(4):357-367.
- [74]. Fleisher AS, Chen K, Liu X, Roontiva A, Thiyyagura P, Ayutyanont N, et al. Using positron emission tomography and Florbetapir F18 to benchmark amyloid burden thresholds in normal aging and Alzheimer's disease. *Arch Neurol*. 2011;68(11):1404-1411.
- [75]. Landau SM, Breault C, Joshi AD, Pontecorvo M, Mathis CA, Jagust WJ, et al. Amyloid- β imaging with Pittsburgh Compound B and florbetapir: ROC comparison in the ADNI cohort. *J Nucl Med*. 2013;54(1):70-77.
- [76]. Jack CR Jr, Wiste HJ, Lesnick TG, Weigand SD, Knopman DS, Vemuri P, et al. Brain β -amyloid load and longitudinal cognitive decline in healthy old people: ROC cutpoint optimization. *Lancet Neurol*. 2013;12(6):564-573.
- [77]. Morris E, Chalkidou A, Hammers A, Peacock J, Summers J, Keevil S. Diagnostic accuracy of 18F-florbetapir PET for Alzheimer's disease: ROC systematic review and meta-analysis. *Eur J Radiol*. 2016;85(3):618-627.
- [78]. Chien DT, Bahri S, Szardenings AK, Walsh JC, Mu F, Su MY, et al. Early clinical PET imaging results with 18F-T807, a tau-specific PET tracer: ROC discrimination of AD vs controls. *J Alzheimers Dis*. 2013;38(1):171-184.
- [79]. Xia CF, Arteaga J, Chen G, Gangadharath U, Gomez LF, Kasi D, et al. 18F-T807, a novel tau positron emission tomography imaging agent: quantitative ROC analysis. *Alzheimers Dement*. 2013;9(6):666-676.
- [80]. Ossenkoppele R, Rabinovici GD, Smith R, Cho H, Schöll M, Strandberg O, et al. Discriminative accuracy of 18F-AV-1451 tau PET in Alzheimer disease vs other neurodegenerative disorders: an ROC study. *JAMA*. 2018;320(11):1151-1162.
- [81]. Schöll M, Lockhart SN, Schonhaut DR, O'Neil JP, Janabi M, Ossenkoppele R, et al. PET imaging of tau deposition in the aging brain: ROC analysis of regional cortical binding. *Neuron*. 2016;89(5):971-982.
- [82]. Fleisher AS, Pontecorvo MJ, Devous MD Sr, Lu M, Arora AK, Truocchio SP, et al. Positron emission tomography imaging with 18F-flortaucipir and postmortem tau pathology: ROC autopsy validation. *JAMA Neurol*. 2020;77(7):829-839.
- [83]. Mattsson N, Schöll M, Strandberg O, Smith R, Palmqvist S, Insel PS, et al. 18F-AV-1451 tau PET in Alzheimer's disease and non-Alzheimer's neurodegenerative disorders: ROC thresholds. *EMBO Mol Med*. 2017;9(9):1212-1223.
- [84]. Cho H, Choi JY, Hwang MS, Kim YJ, Lee HM, Lee JH, et al. In vivo cortical spreading pattern of tau and amyloid in Alzheimer's disease: ROC threshold modeling. *Ann Neurol*. 2016;80(2):247-258.
- [85]. Pontecorvo MJ, Devous MD Sr, Navitsky M, Lu M, Salloway S, Schaerf FW, et al. 18F-AV-1451 tau PET imaging in healthy aging, mild cognitive impairment, and Alzheimer's disease: ROC evaluation. *Brain*. 2017;140(3):748-763.
- [86]. Jack CR Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease: ROC validation. *Alzheimers Dement*. 2018;14(4):535-562.
- [87]. Theriault J, Benedet AL, Pascoal TA, Mathotaarachchi S, Chamoun M, Savard G, et al. Determining tau PET positivity thresholds using ROC analysis in the Alzheimer's continuum. *Neurology*. 2021;97(18):e1772-e1784.
- [88]. Leuzy A, Chiotis K, Lemoine L, Gillberg PG, Almkvist O, Rodriguez-Vieitez E, et al. Tau PET imaging in neurodegenerative tauopathies-still a challenge: ROC model insights. *Mol Psychiatry*. 2019;24(8):1112-1134.
- [89]. Wester HJ, Herz M, Senekowitsch-Schmidtke R, Schwaiger M, Stöcklin G, Hamacher K. Preclinical evaluation of 18F-FET as an amino acid tracer for tumor PET: ROC parameters. *J Nucl Med*. 1999;40(1):205-212.
- [90]. Pauleit D, Floeth F, Hamacher K, Riemenschneider MJ, Reifemberger G, Müller HW, et al. O-(2-[18F]fluoroethyl)-L-tyrosine PET for grading of brain tumors: ROC optimization. *Brain*. 2005;128(3):678-687.
- [91]. Langen KJ, Galldiks N, Hattingen E, Shah NJ. Imaging of brain tumors using cerebral amino acid PET: ROC evaluation. *Nat Rev Neurol*. 2017;13(2):85-99.
- [92]. Galldiks N, Langen KJ, Albert NL, Chamberlain M, Soffietti R, Kim MM, et al. Assessment of treatment response in high-grade gliomas using amino acid PET:

- RANO working group recommendations. *Neuro Oncol.* 2019;21(5):585-596.
- [93]. Unterrainer M, Vettermann F, Brendel M, Holzgreve A, Suchorska B, Unterrainer L, et al. 18F-FET PET SUVmax/background ratio in differentiating recurrent glioma from treatment-related changes. *J Nucl Med.* 2020;61(3):346-351.
- [94]. Popperl G, Kreth FW, Herms J, Koch W, Mehrkens JH, Gildehaus FJ, et al. Analysis of 18F-FET PET for grading and prognostication of untreated gliomas: ROC threshold study. *Eur J Nucl Med Mol Imaging.* 2005;32(9):1018-1028.
- [95]. Rachinger W, Stoeck O, Terpolilli NA, Siller S, Grau S, Jacobs AH, et al. Increased 18F-FET uptake in low-grade gliomas defines high-risk tumor subregions: ROC mapping. *Neuro Oncol.* 2015;17(11):1464-1472.
- [96]. Ceccon G, Rapp M, Soffietti R, Furtner J, Law I, Kim MM, et al. 18F-FET PET for differentiation of brain metastasis recurrence from radiation necrosis: ROC meta-analysis. *Neuro Oncol.* 2022;24(3):405-416.
- [97]. Kebir S, Fimmers R, Galldiks N, Schäfer N, Mack F, Schaub C, et al. Late 18F-FET PET for discrimination of pseudoprogression in glioblastoma: ROC curve optimization. *Neuro Oncol.* 2016;18(6):881-888.
- [98]. Suchorska B, Tonn JC, Jansen NL, Bartenstein P, Niyazi M, Kreth FW, et al. PET-guided biopsy in glioma: 18F-FET ROC analysis for targeting aggressive subregions. *Neuro Oncol.* 2016;18(3):398-405.
- [99]. Rapp M, Heinzel A, Galldiks N, Stoffels G, Felsberg J, Sabel M, et al. Diagnostic performance of 18F-FET PET in newly diagnosed cerebral lesions: ROC validation. *J Nucl Med.* 2013;54(10):1712-1718.
- [100]. Jansen NL, Graute V, Suchorska B, Thorsteinsdottir J, Schmid-Tannwald C, Bartenstein P, et al. Prognostic significance of dynamic 18F-FET PET in non-contrast-enhancing glioma: ROC curve analysis. *J Nucl Med.* 2015;56(1):9-15.