

# Horticultural Approach for *Ex-situ* Conservation of Endangered *Mangifera* species

<sup>1</sup>Nischita.P, <sup>2</sup>M.R.Dinesh\* and <sup>2</sup>Manoj Kumar.H.V

<sup>1</sup>Department of Biotechnology, Centre for Post Graduate Studies, Jain University, Jayanagar, Bengaluru-560011, India

<sup>2</sup>ICAR - Indian Institute of Horticultural Research, Hessaraghatta Lake Post, Bengaluru-560089, India

**Abstract**—Mango (*Mangifera indica* L.) is grown as one of the major fruit crops in all the countries of tropical and subtropical Africa, America and Asia pacific. As there is huge export marketing demand for mango the industrial sector depends on few of the commercial cultivars, resulting in narrowing of the genetic base and flourishing pertaining to the loss of wild species by several anthropogenetic ecological impacts. There is record of more than 60 wild *Mangifera* species identified in South East Asia many are now locally rare and are enclosed on the IUCN Red List categorized as *Mangifera andamanica* (near threatened), *Mangifera camptosperma* (near threatened), *Mangifera casturi* (extinct), *Mangifera griffithii* (no entries found), *Mangifera indica* (data deficient), *Mangifera odorata* (data deficient), *Mangifera sylvatica* (least-concern) and *Mangifera zeylanica* (vulnerable), although there are some conservation initiatives the biodiversity of these wild *Mangifera* species continue to descent. The present study was designed to vegetatively propagate *M. andamanica*, *M. camptosperma* and *M. griffithii*, for long-term sustainment in the gene bank. The establishment of these *Mangifera* species through *ex-situ* conservation was successful using current grafting techniques on *Mangifera indica* root stocks (polyembryonic) and ongoing with initiation in DNA characterisation and assessing for the purpose of breeding.

**Keywords**—Andaman; Conservation; Biodiversity; Extinction; Species; Hotspots.

## I. INTRODUCTION

The forest genetic resources are on major decay and their future in several regions around looks to be disrupted [1,2]. The Convention of biological diversity is solely answerable for conserving, sustaining and employing biodiversity [3,4]. Presently, 34 biodiversity hotspots have been recognized all over the world, occurring in tropical forest. In India, valuable documentation of genetic resource and bioprospecting with remarkable population of extensive flora and fauna exhibits some of the world's most diverse regions viz., Indo-Burma, Himalayas, Andamans Islands and Western Ghats. The genus *Mangifera* is one of the 68 genera originated from Indo-Burma region [5], during the Eocene or an earlier period in the Cretac-

-eous [6], and is usually termed as mango belonging to the family Anacardiaceae. The assessment of conservation rating was undertaken exploiting the International Union for Conservation of Nature (IUCN) released in 1994 at species level [7], where criteria and categories has been evaluated and enhanced numerous times [8]. The Andaman and Nicobar Island of India is a Genetically Diverse Hot Spot (GDHS) dwelling wild mangoes and harbours 25,000 angiosperms, 245 among them are endemic [9]. The native *Mangifera* species in this region are *Mangifera andamanica*, *Mangifera camptosperma*, *Mangifera griffithii*, *Mangifera indica* and *Mangifera nicobarica* [10], enhanced knowledge of the status of such species is necessary for livelihood security and conservation of these valuable species. There is record of more than 60 wild *Mangifera* species identified in South East Asia, many are now locally rare and are categorized on the IUCN Red List as *Mangifera andamanica* (near threatened), *Mangifera camptosperma* (near threatened), *Mangifera casturi* (extinct), *Mangifera griffithii* (no entries found), *Mangifera indica* (data deficient), *Mangifera odorata* (data deficient), *Mangifera sylvatica* (least-concern) and *Mangifera zeylanica* (vulnerable), despite of some conservation projects the biodiversity of these wild *Mangifera* species remain declined. Therefore it is essential to fund collection, *in-situ* and *ex-situ* conservation of these wild species as they are precious resource of biotic and abiotic stress and reinforced 'trait' specific towards crop improvement and assuring against genetic erosion. In this study, we have illustrated the collection and *ex-situ* conservation of *Mangifera* species viz., *Mangifera andamanica*, *Mangifera camptosperma* and *Mangifera griffithii* from Andamans that are locally rare and are included on the IUCN Red List, for further establishment of these species in the gene bank as it is economically important in breeding programmes.

## II. MATERIALS AND METHODS

### A. Study Area

The geographical location of the Andaman and Nicobar Island is 11°42' 59" N – 92° 44' 02" E (Table 1). The highest peak is located in North Andaman Island Saddle Peak at 732m (2,402 ft). It has 572 islands of which 38 are permanently inhabited spreads in an area of 8,249 km<sup>2</sup>. The average annual rainfall as recorded during the year 2007 is 2779.5 mm in Andaman Islands and 2437 mm in Nicobar

\*Corresponding author

M. R Dinesh

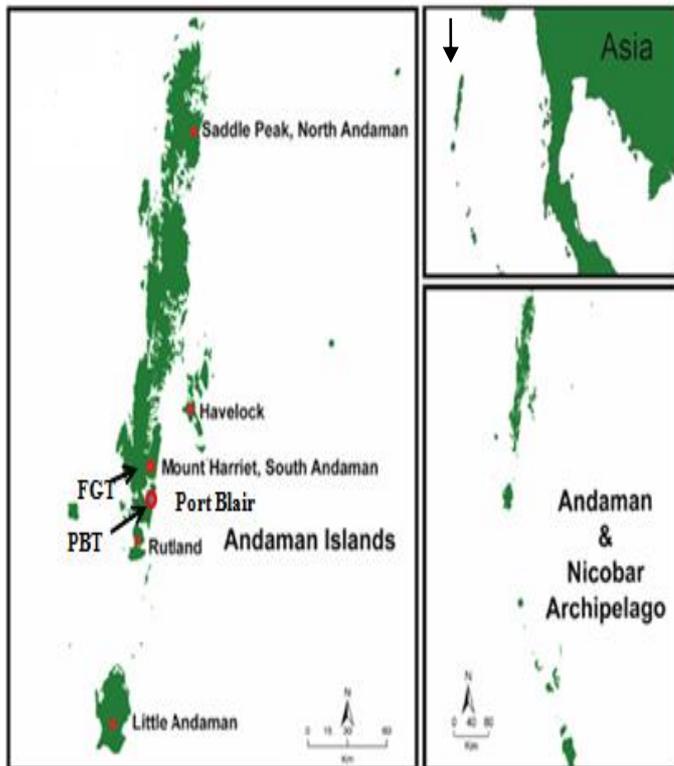
ICAR-IIHR, Hessaraghatta, Lake Post, Bengaluru -89, India

E-mail: [drmrndinesh@gmail.com](mailto:drmrndinesh@gmail.com)

Islands. The vegetation is mostly wet evergreen (North Andaman, South Andaman) and deciduous (Middle Andaman) [11]. The district of South Andamans comprises three different administrative subunits called Tehsils, viz. Port Blair Tehsil (PBT), Ferrargunj Tehsil (FGT) and Little Andaman Tehsil (LAT). The study was undertaken in Chidiyatapu Port Blair Tehsil (PBT) and Chauldari, Jirkhatang, Sholbay, Mt. Harriet Ferrargunj Tehsil (FGT) (**Figure 1**).

Species	Latitude	Longitude
<i>M. andamanica</i>	11° 40" N	92° 43" E
<i>M. andamanica</i>	11° 37" N	92° 33" E
<i>M. camptosperma</i>	11° 40" N	92° 43" E
<i>M. griffithii</i>	11° 43" N	92° 45" E
<i>M. griffithii</i>	11° 67" N	92° 76" E
<i>M. griffithii</i>	11° 40" N	92° 43" E

**Table 1.** The geographical co-ordinates of *Mangifera* species collected from South Andaman Island



**Figure 1.** Map showing the sampling locations for *Mangifera* species surveys covering Port Blair and Ferrargunj Tehsil of South Andaman Island

**B. Collection and propagation of *Mangifera* species**

Methods of *ex-situ* conservation are now accessible and can easily be grouped accordant to the part of the plant that is to be conserved—the whole organism, seed, tissues, or genetic material in culture. The scions, leaf and fruit of *Mangifera* species namely *M. andamanica*, *M. camptosperma*, *M. griffithii* were collected from South Andamans Island (PBT and FGT) during 2014 (**Table 2.**) and transferred to ICAR-IIHR,

Bengaluru. The scions were grafted [12-15], leaves and fruit was utilized for further analysis [16-19].

Species	Tehsil	Place	Location
<i>M. andamanica</i>	FGT	Chauldari	Andaman, India
<i>M. andamanica</i>	FGT	Jirkhatang	Andaman, India
<i>M. camptosperma</i>	FGT	Jirkhatang	Andaman, India
<i>M. griffithii</i>	FGT	Mt. Harriet	Andaman, India
<i>M. griffithii</i>	FGT	Sholbay	Andaman, India
<i>M. griffithii</i>	PBT	Chidiyatapu	Andaman, India

\*FGT - Ferrargunj Tehsil; PBT –Port Blair Tehsil

**Table 2.** The geographical locations of *Mangifera* species collected from South Andaman Island

**III. RESULTS AND DISCUSSION**

In tropical rainforests wild fruit tree species forms an excellent genetic diversity, as they are concerned to wide array of families, genre and subspecies being engaged along with their scope of intraspecies variance [20]. The wild fruit crop of tropical forests considerably imparts to the diversity of species towards nutrition in most of the developing countries [21], but the consumption of the these native fruit are at a major threat due to over exploitation, deforestation and population growth resulting in adverse affect in accessibility of these fruits. There is a wide genetic base in the *Mangifera* species collected from Andaman Island. Currently the grafting an *ex-situ* technique is utilized in tree fruit species viz., almond, apple, apricot, cherry, citrus, grape, mango, peach and plum [22]. In fruit plants the usage of rootstocks has an impact on various parameters like fruit characters, stress factors (biotic & abiotic), yield and ecological conditions by stimulating the scions in improving uptake of nutrients, transporting and utilizing efficiently [23]. The zygotic (monoembryonic) and nuclear (polyembryonic) seedlings are both used as rootstock for grafting [24], monoembryonic seedling shows more inhibition of growth as compared to polyembryonic seedling [25] but use of polyembryonic seedling as rootstock would assure uniformness in fruit orchard unlike zygotic seedlings. Nucellar seedlings are identical to the parent plant [26-28]. However, differences might occur because of somatic variations [29]. Polyembryonic varieties viz., ‘Olour’ have been observed to be moderately tolerant to salt stress [30]. The scions of these wild *Mangifera* species collected were grafted [31] to *M.indica* variety ‘Olour’ under controlled conditions of green house at ICAR-IIHR, Bengaluru, India (**Figure 2**). Successful grafts from three species *M. andamanica* (**Figure 3a and Figure 3b**), *M. camptosperma* (**Figure 3c and Figure 3d**), and *M. griffithii* (**Figure 3e and Figure 3f**) obtained was recorded (**Table 3.**). Seedlings of *M. camptosperma* showed more vigour, success and survival after grafting on *M. indica* rootstock. *Ex-situ* conservation of these wild species and *M. andamanica* (**Figure 3b**) in particular was carried out by collecting of stones and rising of seedlings for further improvement and exploitation. The identification, morphological and molecular characterization of these species was analysed previously [6]. Currently evaluation

of desirable interstocks is implemented to develop these species and genetically characterized as a part of divergence studies within genic collection at ICAR-IIHR, Bengaluru.



Figure 2. ICAR-IIHR Green house in Field Gene Bank

Species	Total Scions	Scions Grafted	Successful Grafts
<i>M. andamanica</i>	15	10	1
<i>M. andamanica</i>	12	10	1
<i>M. camptosperma</i>	10	10	2
<i>M. griffithii</i>	11	06	0
<i>M. griffithii</i>	17	05	0
<i>M. griffithii</i>	12	05	1

Table 3. List of the graft success in *Mangifera* species

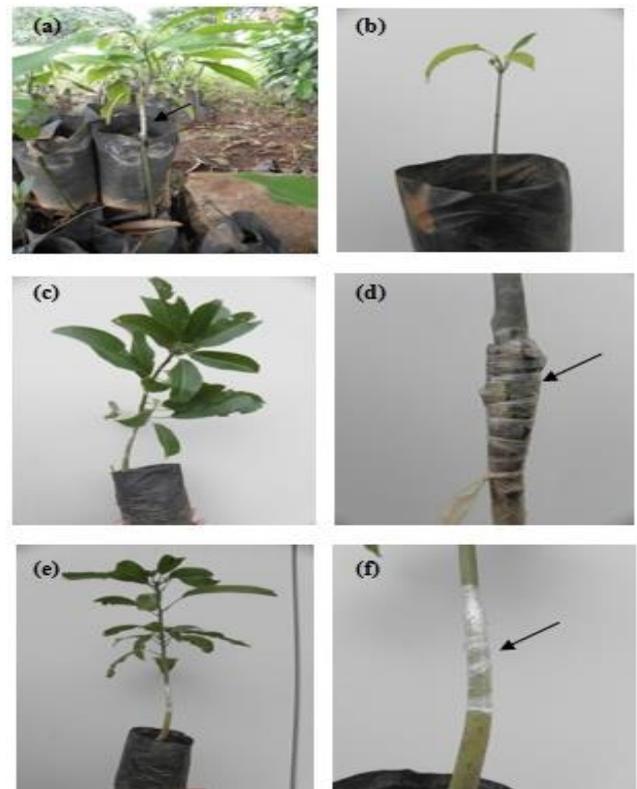


Figure 3. The grafted *Mangifera* species from South Andaman Island

#### IV. FUTURE PROPOSED GUIDELINES

Plants play a major role of Medicare and Agriculture in India representing a natural resource. The above emphasized wild *Mangifera* species are used by tribals and local people to heal several diseases. Therefore, there is huge demand for conservation of these diverse species for the existing and forthcoming generations, by implementing latest technological methods of conservation. *Ex-situ* and *In-situ* conservation of such IUCN red listed species would be excellent step stone towards conservation by discovery of hotspots and collection of wild species more importantly to propagate and conserve redundantly integrating into the breeding projects resulting in the development of the novel crop on the basis of national and international scope. Thus conservation of the species naturally and regenerating artificially is going to be finest choice to retrieve the species from near extinction.

#### ACKNOWLEDGEMENT

The authors thank the Director, IIHR for providing the facilities and the RKVY project for funding the study.

#### Conflict of interest

The authors wish to declare that they have no conflicts of interest.

## REFERENCES

- [1] [Anonymous, “State of the World’s Forests. Food and Agricultural Organization of the United Nations”, Rome, Italy, 1999.
- [2] Anonymous., “Technical guidelines for the establishment and management of ex situ conservation stand of tropical timber species”. International Timber Trade Organization, Yokohama, Japan and Regional Centre for Forest Management, Kuala Lumpur, Malaysia, 2000.
- [3] Anonymous, “UNEP, United Nations Environmental Program. Convention Biological Diversity”, UNEP/BIO/Conf/L, 1992.
- [4] Anonymous., “Millennium Ecosystem Assessment .In: Ecosystems and Human Well Being; Biodiversity Synthesis”, World Resources Institute, Washington, 2005, 85p.
- [5] Hyam R. and Pankhurst R, “Plants and Their Names: A Concise Dictionary” Oxford University Press, Oxford, 1995.
- [6] M .R. Dinesh, K.V. Ravishankar, P.Nischita, B.S.Sandya, B. Padmakar, S. Ganeshan, R. Chithirachelvan and T. V. R. S. Sharma. “Exploration, Characterization and Phylogenetic Studies in Wild *Mangifera indica* Relatives”. Ame. J. Plant .Sci, 2015, 6, 2151-2160.
- [7] IUCN , IUCN Red, “List categories. IUCN Species Survival Commission”. Gland (Switzerland) and Cambridge: IUCN, 1994.
- [8] IUCN ,IUCN Red, “List categories and criteria: version 3.1. IUCN Species Survival Commission” Gland (Switzerland) and Cambridge: IUCN, 2001.
- [9] S.P.S Ahlawat. “Biodiversity of Andaman and Nicobar Islands”. J. An. Sci., 2001,17:1-2.
- [10] S.K Mukherjee, “A monograph on the genus *Mangifera* Linn. Lloydia”, 1949 12: 73-136.
- [11] “Andaman Islands rain forests”. Terrestrial Ecoregions. World Wildlife Fund. Retrieved 28 December 2011.
- [12] R .Kashyap, S.S. Srivastava and A.B. Sharma,, “Studies on the vegetative propagation of mango”. Acta Hort., 1989,231: 263-265.
- [13] P.K Majumder, “Recent advances in propagation and rootstock research in mango-world situation”. Acta Hort., 1988, 213: 157-163.
- [14] S.K. Mukherjee, and P.K. Majumder, Veneer Grafting in Mango. Directorate of Extension, Ministry of Food and Agriculture, New Delhi, India, 1962, pp: 65-66.
- [15] S.K. Mukherjee. and P.K. Majumder, “Effect of different factors on the success in different methods of mango propagation”. Ind. J. Hort., 1964,36: 135-139.
- [16] L. Gielly, and P.Taberlet, “The Use of Chloroplast DNA to Resolve Plant Phylogenies: Noncoding versus rbcL Sequences”. Mol. Bio.Evo., 1994,11, 769-777.
- [17] C. Löhne, and T. Borsch, “Molecular Evolution and Phylogenetic Utility of the petD Group II Intron: A Case Study in Basal Angiosperms”. Mol. Bio.Evo, 2005 ,22, 317-332. <http://dx.doi.org/10.1093/molbev/msi019>.
- [18] B.Oxelman, M. Lidenand and D.Berglund, “Chloroplast rps16 Intron Phylogeny of the Tribe Sileneae (Caryophyllaceae)”. Plant Syst and Evo, 1997, 206, 393-410. <http://dx.doi.org/10.1007/BF00987959>.
- [19] P.Taberlet, L.Gielly, G.Pautou , and J. Bouvet, “Universal Primers for Amplification of Three Non-Coding Regions of Chloroplast DNA”, Plant Mol Bio, 1991, 17, 1105-1109. <http://dx.doi.org/10.1007/BF00037152>
- [20] J.C .Okafor, “Improving edible species of forest products”. Unasyuva, 165, 1991, (42), 17-23.
- [21] C.Okaforl and A. Lamb, “ Fruit trees: diversity and conservation strategies”, Tree Crops and Tropical Ecology Consultants, Kingsway Road, PO Box 3856, Enugu, Nigeria , Agricultural Research Station, PO Box 197, 7bnom, Sabah, Malaysia.
- [22] A. N , Muhammad, Muhammad .I , Qiusheng .K , Fei. C , Waqar. A , Yuan. H and Zhilong B, “Grafting: A Technique to Modify Ion Accumulation in Horticultural Crops” Front. Plant Sci. 2016, 7:1457. <http://dx doi: 10.3389/fpls.2016.01457>.
- [23] L. G. Albrigo, “Rootstocks affect ‘Valencia’ orange fruit quality and water balance”, Proc. Int. Soc. Citriculture, 1977, 1, 62–65.
- [24] I.S.E, Bally, “*Mangifera indica* (mango) Anacardiaceae (cashew family).Species Profiles for Pacific Island Agroforestry”, 2006, www.traditionaltree.org. Accessed on 20 April, 2009.
- [25] J.S. Gora1 , V.K. Singh , D.K. Sarolia1 , Kamlesh Kumar , Rajkumar and V. Bhati, “Performance of Mango (*Mangifera indica* L.) Monoembryonic and Polyembryonic Seedlings under Salt Stress Condition”. Int.J.Curr.Microbiol.App.Sci (2017) 6(6): 3051-3056.
- [26] C .Xiang, and M.L. Roose , “Frequency and characteristic of nucellar and zygotic seedlings in 12 citrus rootstocks”. Sci. Hort, 1988, 37: 47-49.
- [27] R .Garcia, M.J. Asins, J .Forner, and E.A. Carbonell , “Genetic analysis of apomixes in Citrus and Poncirus by molecular markers”. Theor. Appl. Genet, 1999, 99: 511–518.
- [28] C. Ruiz, M.P. Breto, and M.J Asins, “A quick methodology to identify sexual seedlings in citrus breeding programs using SSR markers” Euphy., 2000 ,112: 89–94.
- [29] H.B. Frost, and R.K. Soost, “Seed Production: Development of Gametes and Embryos”, In: Reuther W, Batchelor, HJ Webber HJ (eds) The citrus industry, vol. II. Division of Agricultural Science, University of California, USA, 1968, pp. 292–320.
- [30] A.K . Dubey, A.K Singh. And M. Srivastav, “Salt Stress Studies in Mango- A Review”. Agric. Rev, 2007,28(1):75-78.
- [31] M.N.Islam M.A.Rahim, and A.M.Farooque, “Standardization of Time and Grafting Techniques In Mango Under Bangladesh Condition”, Asian Journal of Plant Science, 2004, 3(3) : 378-386.