

# Practical field Application for Elicitation Efficacy with Gamma Irradiation, Nano- Selenium- Oxide and / or Saccharomyces Servisiae as Elicitors Mediated Amelioration Biomass, Flavonoids, Rutin and Antioxidant capacity of Tartary Buckwheat

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**Abstract:-** For 3 replicate. Statistical analysis of variance for; grain biomass yield (GBY), rutin leaves content (RLC), total flavonoids content (TFC) and rutin content (RC) for grains as well as total antioxidant capacity (TAOC), revealed that both 3 elicitors (G,S,Y) achieved significant enhancement for these 5 traits aside biotic elicitor surpassed Nano- abiotic elicitor (S) that transient physical elicitor (G). Even more, all combined elicitors, i.e. interactions (GS, GY, SY, GSY). Exhibited synergistic significant increment for all 5 traits. Alongside the highest and best positive effect were performed for (G2S3Y3> S3Y3> G2Y3> G2S3. Over all, the results of this study suggested that Yeas - elicitation integrated with gamma irradiation and for name – selenium oxide could be considered as cogitative – oriental strategy to significant Synergistically enhance biomass grain yield, flavonoids, rutin, and antioxidant activity in TBW.

**Keywords:-** Buckwheat, Tartary buckwheat, Elicitation, Gamma- irradiation, Nano- Selenium Particles, Flavonoids, Ration, Antioxidant capacity, Biotic electors, Abiotic elicitation, Physical elicitor.

## I. INTRODUCTION

Buckwheat, a very important edible medicinal plant, has been praised as one of green food for humans in the 21 century [1]. Over recent years the close connections, food and health has become increasing important for consumers, food industry and scientists. Thus, healthy, added – value foods containing bioactive compounds are constantly entering the market. Among grain corps, buckwheat (*Fagopyrum* spp) is gaining momentum because of its content of such healthy compounds [2]. Buckwheat is a minor crop long to the polygonaceae family that can be considered as sustainable crop for its low impute requirement. It has a short growing span [3], it is produced in almost countries where cereals are grown [4], and also the leading continents producers are Europe and Asia [5]. Buckwheat has wide ecological

adaptability, is grows well in infertile land, in adverse climatic conditions and without the use of pesticides and fertilizers.

Buckwheat has played an important role in diets around world over the last 8000 years, mainly in Eastern Europe and Asia [6]. It is considered an important pseudocereal owing to its medicinal and agricultural value. It has various allopathic [7] and medicinal; anti- hypertensive [8], anti- allergic [9]; antibacterial [9-12], protective capacities on DNA damage [13]. These properties are attributed to various nutrients such as vitamins, minerals, amino acids, fatty acids, dietary fibers, phenolic compounds and flavonoids [12,14]. Additionally, buckwheat gluten – free and thus provide an important alternative nutritious food for people with celiac disease [15]. Also, buckwheat seeds are important source that can be processed into various food products such as tea, noodles and pastries [10, 11,16] and BW- Seed – sprouts [17-20]. Buckwheat – seed sprouts, germination process, is the only food processing which provides a significant increase the bioactive secondary metabolites; polyphenols, flavonoids and flavonols during the seven days of germination. Therefore, sprouts can be considered good candidate ingredients for functional foods to be used for lowering the risk of various diseases and / or for exerting health – promoting effects in addition to its nutritive value [17-21]; Exceedingly grains, hay or silage buckwheat can be fed to cattle, sheeps, pigs, goats and poultry, it features similar quality to millet forage [22] but with higher, concentration of protein, 14-15% [23,24].

Investigation of the chemical composition and active principles of buckwheat revealed that grains were rich in constuents, which could be used in contemporary phototherapy. The most important constituents in buckwheat are flavonoids [25-29], in particularly rutin [8], [30- 32] which exhibit various health benefits [33– 38]. The antioxidant capacity of Tartary buckwheat grains is 3-4 times higher than that common buckwheat (CBW) grains, the rutin content in TBW grains is approximately 100 times that in CBW grains [39]. Additionally, the free phenolic content in TBW is 23 to

45 and 25 to 50 folds higher than those in corn and wheat, respectively [40]. rutin extracted from (TBW) have protective and have possible application for treatment of Alzheimer's disease (AD) [35].

The antioxidant potential of BW seeds is determined mainly by phenolic compounds [43]. BW seed contain low molecular weight phenolic, phenolic acids and flavonoids [28,29]. The predominant phenolic compound is rutin, main flavonoids [41,42]. Rutin contribution in BW seeds play a significant role in the antioxidant capacity of BW seeds [30]. These authors demonstrate a strong correlation between rutin content assayed for seeds of various cultivars / accession and their antioxidant capacity ( $r=0.976$ ). A statistically significant correlation ( $r=0.987$ ) also reported by [43] who compared rutin content with antioxidant of seeds and other aerial part of BW. For whole CBW seeds, the contribution of the rutin in the antioxidant capacity was estimated at 25%. Elicitation, with physical [44], abiotic [45,46], biotic [47-49] elicitor is one of few strategies that commercial application in the improvement of bioactive secondary metabolites production from plants as well as cell and organ cultures [50-52,3,53]. It was, therefore, through of trying to find reliable tool to assure further improvement biomass production and bioactive secondary metabolites, under field and in vitro conditions the present study has also conducted.

## II. MATERIAL AND METHODS.

### ➤ Elicitor

Physical elicitor: Tartary buckwheat (TBW); fagopyrum taticum, grains (8% moisture) were subjected to 0, 20, 25 GY gamma irradiation doses / at 1.5 KGY (G1-3) / h, abiotic elicitor: Nano- selenium – oxide particles (S1-3) solution at 0, 20, 40ppb concentrations with 0.01% (V/V) solution of Tween – 20 as surfactant. Biotic elicitor: saccharomyces cervisiae (Y) dissolved in water of concentration 0,0.1,1.0% (W/V) with 0.01% (V/V) solution of Tween – 20 as surfactant. (Y1-3)

### ➤ Planting

At 10 February (2017) gamma irradiated grains were planted in trays contained soil, sand, beat nixed (1: 1: 1 ratio V/V) subsequently established in greenhouse. After 7 days, seedlings were fertilized with nutritive solution; 1.5gm Zn, 0.49gm Cu, 1.2gm. Fe, 1.29mgB, 0.29 gm. Mo/20L water. Seedlings 3 weeks- age were transplanted to the feed (Sandy soil) in plots 4x7m consisted 10 rows, 20cm apart 7m long and 20Cm interspacing (350plant / plot, i.e.12.5 plant / m<sup>2</sup> to give target plant population 52500 plant / Fadden.(125000 plant / h.).

The layout of the field experiment in split – split plot design were conducted with three replicates (3R). the main plot for gamma ray treatments (3G), Nano- selenium= particle (3S) sub- main plot and yeast (Y) sub- sub main plot, that form 81 plots(3Rx3Gx3NSx3Y). plants were irrigated with brackish shallow well water, 1100ppm and were fertigated at 5

weeks from planting with 1.4: 2 NPK, 450Kg/h., through surface irrigation system.

### ➤ Elicitor application

Plants that were resulted from seeds subjected to G1-3 as physical elicitor, at 8 weeks age, were foliarly sprayed with S1-3 and Y1-3 elicitor.

### ➤ Biometric field traits

Just before flowering, at 1st April, 5 representative plants, at random, from each plot, leaves samples were undertaken for rutin content. Whereas, at 20 June plants for each plot were harvested and were quick dried through exposed to sunlight and open air until reach humidity of 12% for grains. Then grain yield per plot and per Fadden were quantitated and converted to grain yield / h (GBY).

### ➤ Bioactive secondary metabolite evaluation

#### • Extraction

Dry leaves and whole grain sample / plot were extracted by ultrasound assisted extraction (UAE) was used for flavonoid extraction, according to [12], under the following conditions; ethanol concentration 65%, solid: liquid ratio of 1:4 gm/ml, extraction time of 35min. and extraction temperature of 35min and extraction temperature of 65°C.

#### • Grain Total flavonoids contents (GTFC)

An aliquot, ml of phenolic / extract was mixed with 1.4 ml of solution containing 25g/L sodium nitrate and 50g/L aluminum nitrate. After 6min, 5ml of 1M sodium hydrate solution were added up to a total volume of 25ml with 95% ethanol. The solution was mixed and incubated at room temperature for 10min. the absorbance was read at 510 nm spectrophotometry (54) and the flavonoid content was calculated with a calibration curve of rutin and expressed as mg quercetin equivalents per gram d .w. grains (mg QE /g.d.w.) per gam of buckwheat grains.

#### • Rutin content in leaves (RLC) and grains rotin content (GRC)

HPLC analysis was monitored at 320 nm and completed using a kromasil 100-5C18 Colum (250mm x 4.6mm 5 μm). Two mobile phase were prepared for mixing: The ratios of methanol: water : acetic acid for mobile phase A and B were (V/V/V) 5: 92.5: : 2.5 and 95: 2.5: 5, respectively. The column was maintained at 35°C and the flow rate was set at 1.0mL min<sup>-1</sup>. The injection volume was 20μL. flavonoids were identified by comparing the retention time with those standard compounds and the peak area was calculated to assess the content of rutin (mg/g d .w).

### ➤ Total antioxidant capacity (TAOC):

The DPPH radical scavenging activity was tested by the method of [55] Briefly, various treatments under investigation,. DPPH solution was also prepared by dissolving 6.0mg. of DPPH in 100ml methanol then, 1.0ml of from

treatment was added into the test tube containing 2.0ml of DPPH solution control was prepared by adding 1.0 of methanol to 2ml of DPPH solution. The mixture was shaken vigorously and was left to stand in the dark for 30min. the absorbance of the resulting solution was measured spectrophotometrically at 517nm. The scavenging activity of DPPH radical was calculated using the following equation: Scavenging activity (%) =  $(1 - (A_{\text{sample}} / A_{\text{control}})) \times 100$

➤ *Statistical analysis:*

The obtained data were subjected to computerized analysis of variance using M- state program. The differences between means were statically tested by the calculated LSD at 1% level.

### III. RESULTS

Statistical analysis of variance revealed that both physical (G), Nano – abiotic (S) and biotic (Y) elicitor as well as their interaction (i.e. combinations); GS, GY, SY, GSY; affected significantly on biomass grains yield /h (BGY), leaves rutin content (RLC), grain total flavonoids content (GTFC), rutin – grain content (GRC) rutin leaves content (RLC) and total antioxidant activity whereas the differences between mean treatment were tested according the calculated LSD at 1% level.

➤ *Biomass grain yield, Kg/h. (BGY)*

(G20, 25Gy), (S20, 40ppb), (Y0.1, 1.0 g/L) achieved significant increment (BGY) as percent of control (100) up to (112,110) (121,126), (125,130) respectively (Table 1), that denote (Y) exceed (S) which excel (G) elicitor. whereas, GS, GY, SY and GSY performed significant synergistic enhancement up to (as % of control), 134,137,142,145, for (G20Gy S40ppb), (G20Gy Y1.0g/L), (S40ppb/ Y1.0 g/L), (G20YG S40ppb ) respectively as represented in Table (1).and represented in Figure (1) .

➤ *Bioactive compound:*

• *Grain total flavonoids (GTFC):*

(G20GY, G25Gy), (S20ppb, S40ppb), (Y0.1g/L, Y 1.0g/L) led to significant enhancement (GTFC), as % of control up to resulted in (135,120), (1.72, 186), (210, 230), respectively. Whilst; (1.72, 186), (186,190), (210, 230), whereas; GS, GY, SY, GSY led to significant synergistic increment, as % of control: up to 225, 270,290,370 for (G20Gy S40ppb), (G20Gy Y0.1g/L), (S1.0g/L Y1.0g/L), respectively, as listed in Table (2)and Figure(2).

• *Bioactive secondary metabolites:*

Rutin leaves content (RLC):(G20Gy, G25Gy), (S20ppb, S40ppb), (Y0.1g/L, Y1.0g/L) increased RLC up to , as % of control (123,117), (135, 147), (142, 160) respectively (Table 2), that also declared (Y) elicitor surpassed (S) elicitor which exceed (G) elicitor, GS, GY, SY, GSY led to significant

synergistic increase (as % of control) up to 173, 182, 186, 200 for ( G20Gy, S40ppb), (G20Gy, Y1.0g/L), (S40ppb y1.0g/L), (G20Gy S40ppb y1.09/L), respectively (Table 3and Figure 3).

• *Grains rutin content (GRC):*

GRC at G1, S1, Y1 as control was 1.052% increased significantly in response to (G20, G25), (S20 S40), (Y0.1, Y1.0) applications up to (180, 162) (205, 230), (220, 255) percent of control, respectively. Combined; i.e interactions, GS, GY, SY, GSY led to significant synergistic increase GRC up to ; 375, 290, 365, 425 as present control for (G20S40), (G20Y1.0), (S40Y1.0), (G20S40 Y1.0) application, respectively (Table 4 and Figure 4).

• *Total antioxidant capacity (TAOC):*

TAOC for Buckwheat extract for control was 60%, while (G20GY G25G0) (S20,40ppb), (Y0.1,1.0g/l) exceed significantly up to (80%, 70%), (85%, 95%), (95%, 115%), respectively Table (5) Figure (5). Elicitor combination, GS, GY, SY, GSY achieved synergistic increase TAOC up to 130, 145, 150, 170 for combination application treatment (G20Gy S40ppb), (G20GyY1.0g/l), (S40ppb Y1.0g/L),( G20Gy S40ppbY1.0g/L), respectively as showed Table 5 and represented Figure (5).

### IV. DISCUSSION

The results indicated that physical (G) elicitor, Nano abiotic elicitors and biotic (Y) elicitor achieved significant enhancement BGY/h, GTFC, RLC, GRC, TAOC aside (Y) surpassed (S) which excel (G). In combined applications (i.e. interactions), GS,GY, SY, GSY resulted significant synergistic enhancement for these five traits, this were attributed to G,S,Y have beneficial effects on morphological physiological and biochemical characters of plant that led to stimulate growth, upraise biomass yield and enhanced formation and accumulation of bioactive compounds as it has been reported for (G) elicitor [56- 58], (S) elicitor [58- 61],(Y) elicitor [61]. Strong radical scavenging activity due to resultant increment of TAOC and RC, were in agreement with has been investigated previously [62].

These findings were in accordance with that has been reported for G [56-58], [59-61] and Y [52]. Whereas, strong radical scavenging activity was due to resultant increment of TAOC and RC were in agreement with that has been investigated previously [62]. Aside, biotic and abiotic elicitors exert their effects by taking part, either directly or in indirectly in multiple signaling pathways by activating transcription genes involved in biosynthesis for Secondary metabolites through up – regulation or downer – regulation) gene expression[63-68]. Further, elicitors can have beneficial effects on morphological, physiological, biochemical characteristics, enhanced growth, biomass production and quality of the formed secondary metabolites [69-75].

**V. CONCLUSION**

Overall the results suggested that elicitation under field application, with G, S and / or Y seems to be reliable powerful technological tool to ameliorate GBY, GTFC, RLC, GRC for TBW. Furthermore, results, also, suggested that the expected

Hyperproduction of rutin extracted from leaves owing to high significant rutin content in leaves that were taken from elicited plants, that required for rutin pharmaceutical industries.

G, S, Y.										
G,S,Y	1		2		3			LSD 1%		
G	1220.2	(100)	1366.2	(112)	1344.7	(110)	12.5			
S	1215.3	(100)	1470.1	(121)	1531.4	(126)	18.2			
Y	1222.7	(100)	1528.4	(125)	1589.5	(130)	21.7			
GS										
G/S	S <sub>1</sub>		S <sub>2</sub>		S <sub>3</sub>			LSD 1%		
G1	1212.4	(100)	1393.2	(115)	1370.5	(113)	23.2			
G2	1370.6	(113)	1539.4	(127)	1624.3	(134)				
G3	1358.9	(112)	1576.6	(139)	1564.8	(129)				
GY										
Gyy	y <sub>1</sub>		y <sub>2</sub>		y <sub>3</sub>			LSD1%		
G1	1219.4	(100)	1463.5	(120)	1438.9	(118)	24.4			
G2	1390.3	(114)	1609.7	(132)	1670.1	(137)				
G3	1414.7	(116)	1646.2	(135)	1585.9	(130)				
Sy										
S/y	y <sub>1</sub>		y <sub>2</sub>		y <sub>3</sub>			LSD 1%		
S1	1209.2	(100)	1529.5	(126)	1487.6	(123)	26.6			
S2	1450.6	(120)	1632.3	(135)	1668.6	(138)				
S3	1511.8	(125)	1693.7	(140)	1717.8	(142)				
GSY										
GSY	S1			S2			S3			LSD 1%
	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	
G1	1217.	1545.	1570.	1223.	1577.	1602.	1219.	1646.	1683.	9.7
	1	7	1	2	9	4	6	5	0	
G2	(100)	(127)	(129)	(100)	(129)	(131)	(100)	(135)	(138)	
	1375.	1606.	1646.	1382.	1675.	1712.	1378.	1695.	1768.	
G3	3	6	1	2	8	5	2	2	4	
	(113)	(132)	(135)	(113)	(137)	(140)	(113)	(139)	(145)	
G3	1363.	1655.	1667.	1369.	1675.	1700.	1365.	1707.	1719.	
	3	3	4	9	8	2	9	4	6	
	(112)	(136)	(137)	(112)	(137)	(139)	(112)	(140)	(141)	

Table 1:- BGY,Kg/h in response to G,S,Y elicitors and their interactions

Gamma ray doses, G<sub>1-3</sub>(0, 20,25GY)-Selenium concentration, S<sub>1-3</sub>(0, 20,40ppb)-Yeast concentration, Y<sub>1-3</sub> (0,0.1,1.0 g/L.) Values between parentheses were percent of control.

G, S, Y.										
G,S,Y	1			2			3			LSD 1%
G	1.23	(100)		1.66	(135)		1.48	(120)		0.09
S	1.31	(100)		2.25	(172)		2.45	(186)		0.10
Y	1.35	(100)		2.86	(210)		3.11	(230)		0.12
GS										
G/S	S <sub>1</sub>			S <sub>2</sub>			S <sub>3</sub>			LSD 1%
G1	1.42	(100)		2.64	(186)		2.70	(190)		0.07
G2	1.96	(138)		3.12	(220)		3.20	(225)		
G3	1.78	(125)		3.05	(215)		3.00	(211)		
GY										
Gyy	y <sub>1</sub>			y <sub>2</sub>			y <sub>3</sub>			LSD1%
G1	1.56	(100)		2.89	(185)		3.06	(196)		0.08
G2	2.25	(144)		3.85	(247)		4.21	(270)		
G3	2.11	(135)		4.14	(265)		3.98	(255)		
Sy										
S/y	y <sub>1</sub>			y <sub>2</sub>			y <sub>3</sub>			LSD 1%
S1	1.51	(100)		3.55	(235)		3.32	(220)		0.11
S2	9.73	(181)		4.15	(275)		4.12	(273)		
S3	2.94	(195)		4.30	(285)		4.38	(290)		
GSY										
GSY	S1			S2			S3			LSD 1%
	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	
G1	1.55	2.38	2.45	1.65	2.66	2.78	1.53	2.59	26.8	0.09
	(100)	(127)	(129)	(100)	(129)	(131)	(100)	(135)	(138)	
G2	2.19	3.02	3.10	2.32	3.98	5.20	2.17	5.43	5.66	
	(141)	(195)	(200)	(145)	(249)	(325)	(142)	(355)	(370)	
G3	1.97	2.82	3.08	2.10	4.08	4.56	2.00	4.06	4.21	
	(127)	(182)	(199)	(131)	(255)	(285)	(131)	(265)	(275)	

Table 2:- TFC,g.QE/100g.d.w.TBWgrains in response to G,S,Y elicitors and their interaction

Gamma ray doses, G<sub>1-3</sub>(0,20,25<sub>Gy</sub>)-Selenium concentration, S<sub>1-3</sub>(0,20,40<sub>ppb</sub>)-Yeast concentration, Y<sub>1-3</sub> (0,0.1,1.0 g/L.)  
 Values between parenthesis were percent of control.

G, S, Y.										
G,S,Y	1			2			3			LSD 1%
G	3.15	(100)		3.88	(123)		3.69	(117)		0.03
S	3.16	(100)		4.27	(135)		4.65	(147)		0.04
Y	3.16	(100)		4.49	(142)		5.06	(160)		0.05
GS										
G/S	S <sub>1</sub>			S <sub>2</sub>			S <sub>3</sub>			LSD 1%
G1	3.17	(100)		4.91	(155)		5.14	(162)		0.04
G2	4.31	(136)		5.17	(163)		5.48	(173)		
G3	3.74	(118)		4.79	(151)		4.69	(148)		
GY										
Gyy	y <sub>1</sub>			y <sub>2</sub>			y <sub>3</sub>			LSD1%
G1	3.18	(100)		5.12	(161)		5.41	(170)		0.05
G2	4.93	(155)		5.44	(171)		5.79	(182)		
G3	4.52	(142)		4.18	(151)		5.25	(165)		
Sy										
S/y	y <sub>1</sub>			y <sub>2</sub>			y <sub>3</sub>			LSD 1%
S1	3.20	(100)		5.12	(160)		5.38	(168)		0.06
S2	5.09	(159)		5.33	(165)		5.92	(172)		
S3	5.28	(165)		5.44	(170)		5.95	(186)		
GSY										
GSY	S1			S2			S3			LSD 1%
	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	
G1	3.21	3.30	4.53	3.20	4.67	5.15	3.22	5.09	5.67	0.03
	(100)	(134)	(141)	(100)	(146)	(161)	(100)	(158)	(176)	
G2	4.01	4.66	5.04	4.06	5.38	5.60	4.06	6.12	6.44	
	(125)	(145)	(157)	(127)	(168)	(175)	(126)	(190)	(200)	
G3	3.82	4.24	4.53	3.84	4.86	5.12	3.90	5.51	5.96	
	(119)	(132)	(141)	(120)	(152)	(160)	(121)	(171)	(185)	

Table 3:- RL,g/100 g.&.w in response to G,S,Y elicitors and their interaction.

Gamma ray doses, G<sub>1-3</sub>(0,20,25<sub>Gy</sub>)-Selenium concentration, S<sub>1-3</sub>(0,20,40<sub>ppb</sub>)-Yeast concentration, Y<sub>1-3</sub> (0,0.1,1.0 g/L.)  
 Values between parenthesis were percent of control.

G, S, Y.										
G,S,Y	1		2			3			LSD 1%	
G	1.052	(100)	1.894	(180)	1.704	(162)			0.08	
S	1.052	(100)	2.157	(205)	2.420	(230)			0.09	
Y	1.052	(100)	2.314	(220)	2.683	(255)			0.11	
GS										
G/S	S <sub>1</sub>		S <sub>2</sub>			S <sub>3</sub>			LSD 1%	
G1	1.065	(100)	2.556	(240)	2.396	(225)			0.05	
G2	2.450	(230)	2.790	(262)	2.929	(275)				
G3	2.364	(222)	2.716	(255)	2.780	(261)				
GY										
Gyy	y <sub>1</sub>		y <sub>2</sub>			y <sub>3</sub>			LSD1%	
G1	1.071	(100)	2.699	(252)	2.870	(268)			0.06	
G2	2.581	(241)	2.945	(275)	3.106	(290)				
G3	2.517	(235)	2.870	(268)	2.945	(275)				
Sy										
S/y	y <sub>1</sub>		y <sub>2</sub>			y <sub>3</sub>			LSD 1%	
S1	1.062	(100)	2.942	(277)	3.069	(289)			0.08	
S2	2.708	(255)	3.133	(295)	3.239	(305)				
S3	2.570	(242)	2.995	(282)	3.876	(365)				
GSY										
GSY	S1			S2			S3			LSD 1%
	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	y <sub>1</sub>	y <sub>2</sub>	y <sub>3</sub>	
G1	1.058	1.852	2.857	1.068	2.777	3.290	1.069	3.742	4.030	
	(100)	(175)	(270)	(100)	(260)	(308)	(100)	(350)	(377)	
G2	1.957	2.325	3.703	1.965	3.898	4.196	1.956	4.272	4.543	
	(185)	(220)	(350)	(184)	(365)	(390)	(183)	(400)	(425)	
G3	1.746	2.190	3.269	1.837	3.759	4.005	1.869	4.009	4.073	
	(165)	(207)	(309)	(172)	(352)	(375)	(175)	(375)	(381)	

Table 4:- GRC % for grains in response to G,S,Y elicitors and their interaction.

Gamma ray doses, G<sub>1-3</sub>(0,20,25<sub>Gy</sub>)-Selenium concentration, S<sub>1-3</sub>(0,20,40<sub>ppb</sub>)-Yeast concentration, Y<sub>1-3</sub> (0,0.1,1.0 g/L.)  
 Values between parenthesis were percent of control.

Application:	TAC%	
Control	60	100
G <sub>2</sub>	80	133
3	70	117
S <sub>2</sub>	85	142
3	95	158
y <sub>2</sub>	95	158
3	115	192
G <sub>2</sub> S <sub>2</sub>	120	200
3	130	217
G <sub>3</sub> S <sub>2</sub>	105	175
3	110	183
G <sub>2</sub> y <sub>2</sub>	130	217
3	145	242
G <sub>3</sub> y <sub>2</sub>	125	208
3	135	225
S <sub>2</sub> y <sub>2</sub>	125	208
3	135	225
S <sub>3</sub> y <sub>2</sub>	140	233
3	150	243
G <sub>2</sub> S <sub>2</sub> y <sub>2</sub>	140	233
3	150	250
G <sub>2</sub> S <sub>3</sub> y <sub>2</sub>	145	242
3	170	283
3	120	200
G <sub>3</sub> S <sub>2</sub> y <sub>2</sub>	120	200
3	130	217
G <sub>3</sub> S <sub>3</sub> y <sub>2</sub>	130	217

Table 5:- TAC for grains extract in response to G,S,Y elicitors and their interaction

Gamma ray doses, G<sub>1-3</sub>(0,20,25<sub>Gy</sub>)-Selenium concentration, S<sub>1-3</sub>(0,20,40<sub>ppb</sub>)-Yeast concentration, Y<sub>1-3</sub> (0,0.1,1.0 g/L.)  
 Values between parenthesis were percent of control.

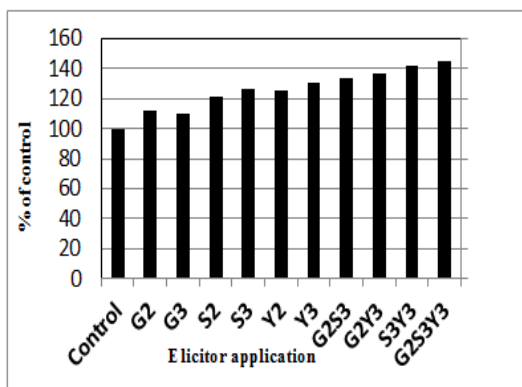


Fig 1:- BGY/h as% of control

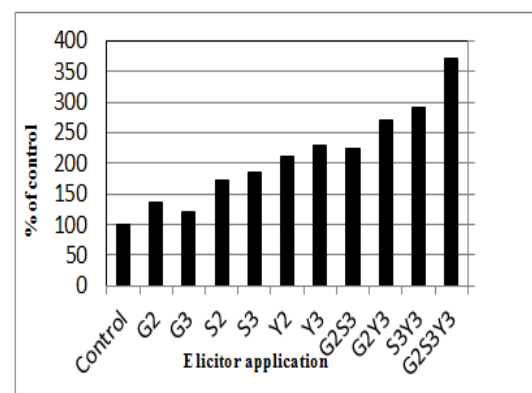


Fig 2:- GTFC as% of control



REFERENCES

- [1]. Wang, C.; She, H. Z.; Liu, X. B.; Hu, D.; Ruan, R. W.; Shao, M. B.; Zhang, L. Y.; Zhou, L. B.; Zhang, G. B.; Wu, D. Q.; Yi, Z. L. Effects of fertilization on leaf photosynthetic characteristics and grain yield in tartary buckwheat Yunqiao1. *Photosynthetica*; 2017. 55(1):77-84.
- [2]. Christa, K., Soral-Šmietana, M., (2008). Buckwheat grains and buckwheat products—nutritional and prophylactic value of their components—a review. *Czech J. Food Sci.* 26, 153–16
- [3]. Zhou ML. Bai DQ. Tang Y. Zhu xm Shao JR (2012). Genetic diversity of our new species related to southwestern Sichuan buckwheats as revealed by karyotype ISSR and allozyme characterization plant *Evol.* 298:751-759.
- [4]. Campbell CG (1979). Buckwheat, *Fagopyum esculentum* Munch promoting the conservation and use of underutilized and neglected crop. 19. Rome. *int. plant Genet. Resour. Institute* 39 p.
- [5]. FAOSTAT (Food and Agricultural Organization of the United Nations Statistics (2013). FAO Statistical Databases Accessed online at <http://faostat.fao.org/>. FAO. Rome.
- [6]. Rana, J.C., Singh, M., Chauhan, R.S., Chahota, R.K, Sharma, T.R., Yadav, R., & Arched S. (2016).
- [7]. Golisz, A.; Lata, B.; Gawronski, S.W.; Fuji, Y. (2007). Specific and total activities of the allelochemicals identified in buckwheat. *Weed Biol. Manag.* 7, 164–171.
- [8]. Kim, S.-J.; Zaidul, I.; Suzuki, T.; Mukasa, Y.; Hashimoto, N.; Takigawa, S.; Noda, T.; Matsuura-Endo, C.; Yamauchi, H. (2008). Comparison of phenolic compositions between common and Tartary buckwheat (*Fagopyrum*) sprouts. *Food Chem.* 110, 814–820.
- [9]. Amarowicz, R.; Dykes, G.A.; Peg, R.B. (2008). Antibacterial activity of tannin constituents from *Phaseolus vulgaris*, *Fagopyrum esculentum*, *Corylus avellana* and *Juglans nigra*. *Fitoterapia.* 79, 217–219.
- [10]. Ma, Y. J., Guo, X. D., Liu, H., XU, B. N., & Wang, M. (2013). Cooking, textural, sensorial, and antioxidant properties of common and tartary buckwheat noodles. *Food Science and Biotechnology*, 22, 153-159.
- [11]. Merendino, N., Molinari, R., Costantini, L., Mazzucato, A., Pucci, A., Bonafaccia, F Bonafaccia, G. (2014). A new functional” pasta containing tartary buckwheat sprouts as an ingredient improves the oxidative status and normalizes some blood pressure parameters in

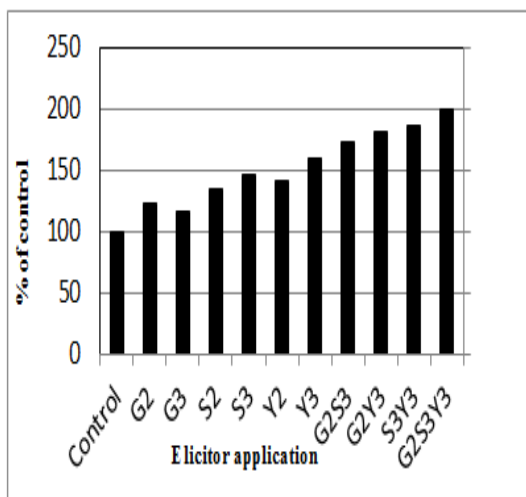


Fig 3:- RLC as % of control

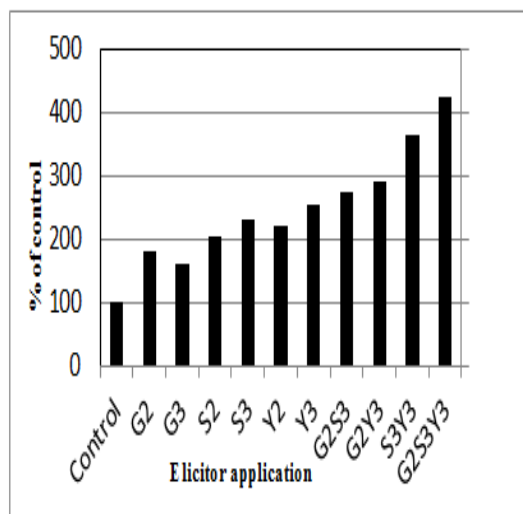


Fig 4:- GRC as % of control

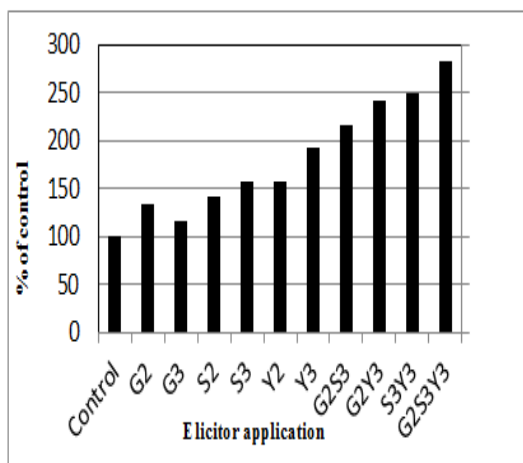


Fig 5:- TAOC as % of control.

- spontaneously hypertensive rats. *Food & Function*, 5, 1017–1026.
- [12]. Zheng, C., Hue, C., Ma, X., Peng, C., Zhang, H., & Qi, L. (2012). Cytotoxic phenylpropanoid glycosides from *Fagopyrum tataricum* (L.) Gaertn. *Food Chemistry*, 132, 433–438.
- [13]. Cao, W., Chen, W. J., Suo, Z. R., & Yao, Y. P. (2008). Protective effects of ethanolic extracts of buckwheat grouts on DNA damage caused by hydroxyl radicals. *Food Research International*, 41, 924–929. [14] Pongrac, P., Vogel-Mikuš, K., Regvar, M., Vavpetic, P., Pelicon, P., & Kraft, I. (2011). Improved lateral discrimination in screening the elemental composition of buckwheat grain by micro-PIXE. *Journal of agricultural and food chemistry*, 59(4), 1275–1280.
- [14]. Pongrac, P., Vogel-Mikuš, K., Regvar, M., Vavpetic, P., Pelicon, P., & Kraft, I. (2011). Improved lateral discrimination in screening the elemental composition of buckwheat grain by micro-PIXE. *Journal of agricultural and food chemistry*, 59(4), 1275–1280.
- [15]. Giménez-Bastida, J.A., Piskula, M., & Zieliński, H. (2015). Recent advances in development of gluten-free buckwheat products. *Trends in Food Science & Technology*, 44, 58–65.
- [16]. Oin, P., Wu, L., Yao, Y., & Ren, G. (2013). Changes in phytochemical compositions, antioxidant and  $\alpha$ -glucosidase inhibitory activities during the processing of tartary buckwheat tea. *Food Research International*, 50, 562–567.
- [17]. Hsu, C. K., Chiang, B. H., Chen, Y. S., Yang, J. H., & Liu, C. L. (2008). Improving the antioxidant activity of buckwheat (*Fagopyrum tataricum* Gaertn) sprout with trace element water. *Food Chemistry*, 108(2), 633–641.
- [18]. Kim, H. J., Park, K. J., & Lim, J. H. (2011). Metabolomic analysis of phenolic compounds in buckwheat (*Fagopyrum esculentum* M.) sprouts treated with methyl jasmonate. *Journal of Agricultural and Food Chemistry*, 59(10), 5707–5713.
- [19]. Lim, J. H., Park, K. J., Kim, B. K., Jeong, J. W., & Kim, H. J. (2012). Effect of salinity stress on phenolic compounds and carotenoids in buckwheat (*Fagopyrum nesculentum* M.) sprout. *Food Chemistry*, 135, 1065–1070.
- [20]. Park, W. T., Kim, Y. B., Seo, J. M., Kim, S. J., Chung, E., Lee, J. H., & Park, S. U. (2013). Accumulation of anthocyanin and associated gene expression in radish sprouts exposed to light and methyl jasmonate. *Journal of Agricultural and Food Chemistry*, 61(17), 4127–4132.
- [21]. Brajdes, C.; Vizireanu, C. (2012). Sprouted buckwheat an important vegetable source of antioxidants. *Annals of the University "Dunarea de Jos" of Galati - Fascicle VI: Food Technology*; 36(1):53-60.
- [22]. Gorgon AV (2013). Produtividade e qualidade da forragem de milho (pennisetumglaucum (L.) R. BR) e de trigo mourisco (*Fagopyrum esculentum*. Moench) cultivado no cerrado. Bacheior in Agronomy Monograph. Universidade Brasilia. Brazil p 49.
- [23]. Sonia Mann, Deepika Gupta, & Rajinder K Gupta. (2012). Evaluation of nutritional and antioxidant potential of Indian Buckwheat grains ingian journal of Traditional Knowledge Vol .11(1) ,pp.40 – 44.
- [24]. Mohammad Reza Sobhani1, Gulahmad Rahmikhdoev, Dariush Mazaheri and Majid MajidianInfluence of different sowing date and planting pattern and N rate on buckwheat yield and its quality Austrajian Journal of Crop Science AJCS 8(10):1402-1414.
- [25]. Kim, J.H.; Beak, M.H.; Chung, B.Y.; Wi, S.G.; J.S. Kim. (2004): Alternations in photo – synthetic pigments and antioxidant machineries of red pepper (*Capsicum annum* L. Seedlings from gamma irradiated seeds. *J. Plant Biol.*; 47:314-321.
- [26]. Lan-Sook Lee, Eun-Ji Choi, Chang-Hee Kim, Jung-Min Sung, Young-Boong Kim, Dong-Ho Seo, Hyun-Wook Choi, Yun-Sang Choi, Jun-Seok Kim, Jong-Dae Park. (2016) .Contribution of flavonoids to the antioxidant properties of common and Tartary buckwheat *Journal of Cereal Science* 68 ,181e186.
- [27]. Zielińska D., Szawara-Nowak D., Michalska A., (2007). Antioxidant capacity of thermally-treated buckwheat. *Pol. J. Food Nutr. Sci.*, 57, 465–470.
- [28]. Verardo V., Arráez-Román D., Segura-Carretero A., Marconi E., Fernández-Gutiérrez A., Caboni M.F. (2010). Identifi cation of buckwheat phenolic compounds by reverse phase high performance liquid chromatography-electrospray ionization-time of fl ightmass spectrometry(RP-HPLC-ESI-TOF-MS). *J. Cereal Sci*, 52, 170–176.
- [29]. Inglett G.E., Chen D., Berhow M., Lee S. ( 2011). Antioxidant activity of commercial buckwheat fl ours and their free and bound phenolic compositions. *Food Chem.* 125, 923–929.
- [30]. Jiang P., Burczynski F., Campbell C., Pierce G., Austria J.A., Briggs C.J. (2007). Rutin and flavonoid contents in three buckwheat species *Fagopyrum esculentum*, *F. tartaricum*, and *F. homotropicum* and their protective

- effects against lipid peroxidation. *Food Res. Int.*, 40, 356–364.
- [31]. Kalinova, J.; Triska, J.; Vrchotova, N. (2006). Distribution of vitamin E, squalene, epicatechin, and rutin in common buckwheat plants (*Fagopyrum esculentum* Moench). *J. Agric. Food Chem.* 54, 5330–5335.
- [32]. Kreft, I.; Fabjan, N.; Yasumoto, K. (2006). Rutin content in buckwheat (*Fagopyrum esculentum* Moench) food materials and products. *Food Chem.* 98, 508–512.
- [33]. Lee Dong GI; Jang Iksoon; Yang kyeongEun; Yoon SoJung; Baek Su Jeong; Lee JooJong;Suzuki, T.;Chung keun Yook; Woo Sunhee; Choi Jong Soon. (2016). Effect of rutin from Tartary buckwheat sprout on serum glucose- lowering in animal model of type 2 diabetes. *Acta pharmaceutica (Zagreb)*. 66(2): 297-202.
- [34]. Brunori,A.; Varga, A; Szedljak, I; Vegvari, G. (2016). Combined effect of heat treatment and humidity on the total polyphenol content of Tartary buckwheat whole flour. *Food Technology*, 40(2): 135-140.
- [35]. Ji Yeon Choi, Jeong Min Lee, Dong Gu Lee, Sunghun Cho, Young-Ho Yoon, Eun Ju Cho, and Sanghyun Lee. (2015). The n-Butanol Fraction and Rutin from Tartary Buckwheat Improve Cognition and Memory in an In Vivo Model of Amyloid-b-Induced Alzheimer's Disease *J Med Food* 18 (6) .631–641 .
- [36]. Ihme N, Kiesewetter H, Jung F, Hoffmann KH, Birk A et al., Leg. (1996) . edema protection from buckwheat herb tea in patients with ehronic venous insufficiency : a single – centre randomized. double blind . placebo – controlled clinical trial . *Eur J Clin pharmacol*, 50 ,443- 447.
- [37]. Pulido R, Bravo L& Saura – Calixo F, (2000) .Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing / antioxidant power assay, *J Agri Food Chem* 48 3396 – 3402.
- [38]. Dong-GI lee1, ik soon jang, kyeong eun yang1, so-jung yoon1, sujeong baek, joo yong le, tatsuro Suzuki, keun-yook chung, sun-he woo, Jong-soon choi1, (2016) Effect of rutin from Tartary buckwheat sprout on serum Glucose-lowering in animal model of type 2 diabetes *Acta Pharm.* 66 ,297–302.
- [39]. Morishita, T., Yamaguchi, H., & Degi, K. (2007). The contribution of polyphenols to antioxidative activity in common buckwheat and Tartary buckwheat grain. *Plant Production Science*, 10(1), 99–104.
- [40]. Guo, X., Zhu, K., Zhang, H., & Yao, H. (2010). Anti-tumor activity of a novel protein obtained from Tartary buckwheat. *International Journal of Molecular Sciences*, 11, 5201–5211.
- [41]. Zielińska D., Turemko M., Kwiatkowski J., Zieliński H., Evaluation of flavonoid contents and antioxidant capacity of the aerial parts of common and Tartary buckwheat plants. *Molecules*, 2012, 17, 9668–9682.
- [42]. Kiprovski B., Mikulic-Petkovsek M., Slatnar A., Veberic R., Stampar F., Malencic D., Latkovic D. (2015). Comparison of phenolic profile and antioxidant properties of European *Fagopyrum esculentum* s cultivars. *Food Chem.*, 185, 41–47.
- [43]. Holasova M., Fiedlerova V., Smrcinova H., Orsak M., Lachman J., Vavreinova S. (2002), Buckwheat – the source of antioxidant activity in functional foods. *Food Res. Inter.*, 35, 207–211.
- [44]. Dörnenburg,H; Knorr, D. (1995).Strategies for the improvement of secondary metabolite production in plant cell cultures. *Enzyme Microb Technol.* 17, 674–684.
- [45]. Suvarnalatha, G.; Rajendran, L.; Ravishankar, G.A.; Venkataraman, L.V. (1994). Elicitation of anthocyanin production in cell cultures of carrot (*Daucus carota* L.) by using elicitors and abiotic stress. *Biotechnol. Lett.* 16, 1275–1280.
- [46]. Radman, R.; Sae, T.; Bucke, C. and T. Keshavarz. (2003):Elicitation of plant and microbial systems *Biotechnol. Appl Biocchem.*37:91-102.
- [47]. Sandra. Pitta–Alvarez, Tatiana C. Spollansky, Ana M. (2000) . Giulietti The influence of different biotic and abiotic elicitors on the production and profile of tropane alkaloids in hairy root cultures of *Brugmansia candida* *Enzyme and Microbial Technology* 26 ,252–258.
- [48]. Randhir, R.; Lin, Y.-T.; Shetty, K. (2004). Stimulation of phenolics, antioxidant and antimicrobial activities in dark germinated mung bean sprouts in response to peptide and phytochemical elicitors. *Process Biochem.* 39, 637–646.
- [49]. Angelova, Z.; Gergiev, S.; Roes, W. (2006). Elicitation of plants. *Biotechnol. l. Equip.* 20, 72–83.
- [50]. Michał Świeca .(2016). elicitation and treatment with precursors of phenolics synthesis improve low-molecular antioxidants and antioxidant capacity of buckwheat sprouts *Acta Sci. Pol. Technol. Aliment.*, 15(1), 17–28. DOI: 10.17306/J.AFS. 1.2.
- [51]. Kiran Sharma, Rasheeduz Zafar .(2016) . Optimization of methyl jasmonate and-cyclodextrin for enhanced duction of taraxerol and taraxasterol in (*Taraxacum officinale* Weber) cultures *Plant Physiology and Biochemistry* 103 ,24e30 .

- [52]. Urszula Zlotek and Michał Swieca. (2016). Elicitation effect of *Saccharomyces cerevisiae* yeast extract on main health-promoting compounds and antioxidant and anti-inflammatory potential of butter lettuce (*Lactuca sativa* L.) JSci Food Agric; 2565-2572.
- [53]. Nieves Baenas, Cristina García-Viguera and Diego A. Moreno. (2014). Elicitation: A Tool for Enriching the Bioactive Composition of Foods *Molecules*. 19, 13541-13563.
- [54]. Xue, C., Yuan, S., Wang, P., Yao, C., & Niu, J. (2006). On the selection of methods for the determination of flavonoids in extracts of spring buckwheat powder. *Physical Testing and Chemical Analysis Part B (Chemical Analysis)*, 42(1), 21–23.
- [55]. Chan EWC, Lim YY, Omar M (2007). Antioxidant and antibacterial activity of leaves of *Etilingera species* (Zingiberaceae) in Peninsular Malaysia. *Food Chem.*, 104(4): 1586-1593.
- [56]. [56] Potrebko, I. and A.V.A. (2009). Resurreccion. Effect of Ultraviolet doses combines ultraviolet – ultrasound treatments on trans – resveratrol and trans – piceid content in sliced peanut kernels. *Journal of Agricultural and Food Chemistry*, 57(17): 7750-7756.
- [57]. Cho, K.; Dea Wook, K.; YoungHo, J.; Shibato, J.; Tamogami, S.; Yonekura, M.; Jwa, N. S.; Kubo, A.; Agrawal, G. K. and R. Rakwal. (2007): Light dark responsiveness of Kinetin-
- [58]. Kiong, A.; Ling, A.; Pick, S. H.; Grace L. and A.R. Harun. (2008). Physiological responses of *Orthosiphon stamineus* plantlets to gamma irradiation. *American Eurasian Journal Sustainable Agriculture*, 2(2): 135-149.
- [59]. Monika. A & Romuald. S & Joanna Kowalska & Grazyna .(2015). Bystrzejska-Piotrowska Accumulation of Platinum Nanoparticles by *Sinapis alba* and *Lepidium sativum* Plants *Water Air Soil Pollut* 226: 126.
- [60]. Hina Fazal Bilal Haider Abase Nisar Ahmad Mohammad Ali (2016). Elicitation of Medicinally Important Antioxidant Secondary Metabolites with Silver and Gold Nanoparticles in Callus Cultures of *Prunella vulgaris* L. *Appl Biochem Biotechnol*, 180:1076–1092.
- [61]. Faezeh Ghanati<sup>1</sup> Somayeh Bakhtarian<sup>1</sup>, Behrooz Mohammad Parast<sup>2</sup> and Mahboobeh Keyhani (2014). Behrooz Production of New Active Phytocompounds by *Achillea millefolium* L. after Elicitation with Silver Nanoparticles and Methyl Jasmonate *Biosciences Biotechnology Research Asia*, 2 Vol. 11(2), 391-399.
- [62]. Da Bing Xiang, Gang Zhao, Yan Wan, Mao Ling Tan, Chao Song & Yue Song (2016). Effect of planting density on lodging-related morphology, lodging rate, and yield of Tartary buckwheat (*Fagopyrum tataricum* Plant Production Science, VOL. 19, NO. 4, 479–488
- [63]. Hina Fazal , Bilal Haider Abbasi , Nisar Ahmad, Mohammad Ali (2016) Elicitation of Medicinally Important Antioxidant Secondary Metabolites with Silver and Gold Nanoparticles in Callus Cultures of *Prunella vulgaris* Lapp *Biochem Biotechnol* 180:1076–1092.
- [64]. Hassn, M.; Bae Han Hong (2017). An overview of stress – induced resveratrol synthesis in grapes: Perspectives for resveratrol-enriched grape products. *Molecules*; 22(2):294.125.
- [65]. Xiaohua Li, Aye Aye Thwe, Chang Ha Park, Sun Ju Kim, Mariadhas Valan Arasu, Naif Abdullah Al-Dhabi, Sook Young Lee & Sang Un Park (2017). Ethephon-induced phenylpropanoid Accumulation and related gene expression in Tartary buckwheat (*Fagopyrum tataricum*(L) Gaertn.) hairy root. *Biotechnology & Biotechnological Equipment* 1314-3530.
- [66]. Zlotek, U. (2017). Effect of jasmonic acid and yeast extract elicitation on low-molecular antioxidants and antioxidant activity of marjoram (*Origanum majorana* L.). *Acta Scientiarum polonorum – Technologia Alimentaria*; 16(4):371-377.
- [67]. Chung Min; Kaliyaperumal Rekha; Govindasamy Rajakumar; Muthu Thiruvengadam (2018). Production of bioactive compounds and gene expression alterations in hairy root cultures of Chinese cabbage elicited by copper oxide nanoparticles. *Tissue and Organ Culture*; 134(1):95-106.
- [68]. Mosadegh, H.; Trivellini, A.; Ferrante, A.; Lucchesini, M.; Vernieri, P.; Mensuali, A. (2018). Applications of UV-B lighting to enhance phenolic accumulation of sweet basil. *Scientia Horticulturae*; 299:107 – 116.
- [69]. Sharifi-Rad<sup>1</sup>, M. Sharifi-Rad<sup>2</sup>, and J. A. (2016). Teixeira da Silva<sup>3</sup> Morphological, Physiological and Biochemical Responses of Crops (*Zea mays* L., *Phaseolus vulgaris* L.), Medicinal Plants (*Hyssopus officinalis* L., *Nigella sativa* L.), and Weeds (*retroflexus Amaranthus* L., *Taraxacum officinal* F. H. Wigg) Exposed to SiO<sub>2</sub> Nanoparticles *J. Agr. Sci. Tech.* Vol. 18: 1027-1040.
- [70]. Jaing Y.; Zeng, Z.H.; Bu, Y.; Ran, C. Z.; Li, J.Z.; Han, J. J.; Tao, C.; Zhang, K.; Wang, X. X.; Lu, G. X.; Li, Y. J.;

- Hu, Y. G. (2015). Effects of selenium fertilizer on grain yield, Se uptake and distribution in common buckwheat (*Fagopyrum esculentum Moench*). *Plant Soil and Environment*. 61(8):371 – 377.
- [71]. Rabia Javed, Muhammad Usman, Buhara Yucsan, Muhammad Zia, Ekrem Gurel (2017). Effect of zinc oxide (ZNO) nanoparticles on physiology and steviol glycosides production in micropropagated shoots of *Stevia rebaudiana* Bertoni. *Plant physiology and Biochemistry*. 110 – 94 – 99.
- [72]. Moharrami, F.; Hosseini, B.; Sharafi, A.; Farjaminezhad, M. (2017). Enhanced production of hyoscyamine and scopolamine from genetically transformed root culture of *Hyoscyamus reticulatus* L. elicited by iron oxide nanoparticles. *In vitro Cellular & Developmental Biology- Plant*; 35(2):104 -111.
- [73]. Nneugart, S.; Schreiner, M. (2018). UVB and UVA as eustressors in horticultural and agricultural crops. *Scientia Horticulture*. 234 : 370 – 381.
- [74]. Gou JunBo; Hao FuHua; Huang Chong Yang; Know MoonHyunk; Chen FangFang; Li ChangFu; Liu Chao Yang; Ro DaeKyun; Tang HuiRu; Zhang YanSheng. (2018). Discovery of a non-stereoselective cytochrome P450 catalyzing either 8 alpha – or 8 beta hydroxylation of germacrene A acid from the Chinese medicinal plant, *Inula hupehensis* plant journal. 93(1):92-106.
- [75]. Fereshte Moharrami . Bahman Hosseini . Ali Sharafi . Manouchehr Farjaminezhad (2017). Enhanced Production of hyoscyamine and scopolamine from genetically transformed root culture of *Hyoscyamus reticulatus* L. elicited by iron oxide nanoparticles. *In Vitro Cell.Dev.Biol.\_ plant*(2017) 53:104\_111.