ISSN No:-2456-2165

# Study and Isolation of Skin Microflora and their Effects on Human Health

Dr Madhulika Singh, Priya Rathore, Shuchi Gupta Department of Biotechnology, Sadhu Vaswani College Bairagarh, Bhopal

Abstract:- Staphylococcus species are Gram positive, cocci shaped bacteria found especially in the wounds of a person. Skin micro flora is the major source of this species and present in the upper epidermis of it. Skin samples were collected to isolate the Staphylococcus bacteria and studied their resistance towards antibiotics. In this thesis, samples of different communities were taken by sterilized swap sticks and inoculate. Mannitol Salt Agar Media specialized for Staphylococcus Species was used to culture the incubated broth tubes to study the morphology. Biochemical test and Antibiotic Sensitivity Tests were performed to analyze the enzyme activity and Sensitivity Resistance towards antibiotics, respectively. This study revealed the two species of Staphylococcus- aureus and epidermidis and their morphology and biochemical characteristics along with resistance towards some antibiotics.

#### **INTRODUCTION**

The skin serves as a natural protective barrier against invasion by most microorganisms because of the acidity of the skin, the presence of indigenous flora, the temperature of the skin sebum secreted by oil glands and creation of hypertonic environment by the salts in perspiration. However perspiration and sebum serve as a source of nutrition for certain micro organism and help in their establishment as normal flora of the skin most encountered bacteria on the skin are gram positive cocci including micrococcus and staphylococcus. Micrococcus is generally considered to be a saprophyte while *staphylococci* which include 40 species, most of them are harmless and reside normally on the skin and mucous membranes of humans and other organisms. Staphylococcus aureus which is a member of the normal flora of the skin is an important and frequently encountered human pathogen. Sebum secreted from oil glands helps the survival of propioniobacterium acnes, an anaerobic gram-positive rod in hair follicles. The so-called normal flora (or biota) of the skin vary according to the skin region and the environmental factors. Most pathogenic organisms do not penetrate the unbroken skin but as soon as epidermis is destroyed infection easily occurs. Staphylococci and certain fungi are able to penetrate the hair follicles and cause disease in the deeper tissues. Although human skin harbours many different genera, this exercise deals with the isolation and identification of catalase-positive, salt tolerant members of gram-positive cocci belonging to the family *Micrococcaceae*(*Actinobacteria*) and *Staphylococcaceae* (*Fimicutes*).

## METHODOLOGY

Chemical; Nutrient Broth - HIMEDIA M001- 500 Gms Mannitol Salt Agar- Microgen – DM1118. Mueller Hinton Agar: - Microgen – DM 1173. BIO- CHEMICALS MEDIA:-Coagulase Mannitol Agar Base-HIMEDIA- M272- 100gm. ANTIBIOTICS:-HEXA G- PLUS 1- HIMEDIA- HX001-1PK

Antibiotics	Concentration	
Penicillin G	10 mcg	
Oxacilliin	1mcg	
Cephalothin	30mcg	
Clindamycin	2mcg	
Erythromycin	15mcg	
Amoxcyclav	30mcg	
TADIE, 1		

TABLE:-1

Antibiotics	Symbol	Concentration
Tetracycline	TE	30mcg
Co- Trimoxazole	СОТ	25mcg
Cloxacillin	COX	1mcg
Lincomycin	L	2mcg
Cefuroxime	CXM	30mcg
Cefotaxime	CTX	30mcg
	TADLE 0	

TABLE:-2

## HEXA G- PLUS 5- HIMEDIA- HX005- 1PK.

Sample collection - From skin of different communities -From railways station sample, Slum area. The study was undertaken in Sadhu Vaswani College, Botany & Biotechnology Department between Feb to May, 2014. (32) Clinical specimens taken by swab from various aged individuals from different communities such as -- with different locations of skin under sterile conditions. The staphylococcus species procedure is determining the identity of Staphylococcus species as below:

Composition of Ingredients-

Ingredients	Gms / Litre
Protease peptone	10 gm
Sodium chloride	75gm
Beef extract	10gm
D-Mannitol	1.5gm
Phenol red	0.025gm
Agar	15gm
Final pH (at 25°C)	$7.4{\pm}0.2$
-	

\*\*Formula adjusted, standardized to suit performance parameters

Composition of Ingredients-

Ingredients	Gms / Litre
Peptic digest of animal tissue	5.0 gm
Sodium chloride	5.0gm
Beef extract	1.5gm
Yeast extract	1.5gm
Agar	15gm
Final pH (at 25°C)	7.4±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### *Growth in Liquid media:*

Nutrient Broth is used for the general cultivation of less fastidious microorganisms, can be enriched with blood or other biological fluids.

Composition of Ingredients-

Ingredients	Gms / Litre
Peptic digest of animal tissue	5.0 gm
Sodium chloride	5.0gm
Beef extract	1.5gm
Yeast extract	1.5gm

\*\*Formula adjusted, standardized to suit performance parameters.

Composition\*\*

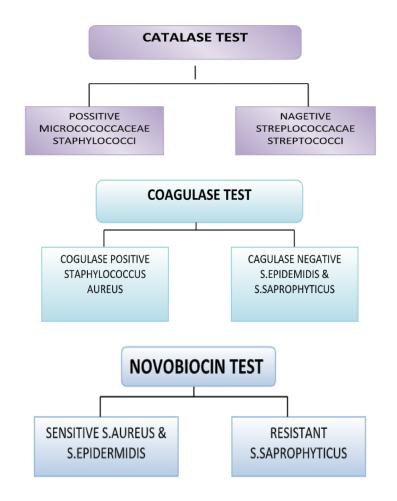
Ingredients	Gms / Litre
Peptic digest of animal tissue	10.000
Sodium chloride	5.000
Final pH (at 25°C)	7.2±0.2

\*\*Formula adjusted, standardized to suit performance parameters

Study of External Morphology by staining procedure:

Gram staining Ingredients	Composition	
Grams Crystal Violet (S012)	Ingredients	Grams/Litre
	Crystal violet	2.0 gm
	Ammonium oxalate	0.8 gm
	Ethyl alcohol 95%	20.0 ml
	Distilled water	80.0 ml
Grams Iodine (S013)	Iodine	1.0 gm
	Potassium iodide	2.0 gm
	Distilled water	300.0 ml
Grams Decolourizer (S032)	Ethyl alcohol (95%)	50 ml
	Acetone	50ml
Safranin (S027), 0.5% w/v (Gram's Counter stain)	Safranin	0.05gm
	Ethyl alcohol 95%	100ml

Biochemical tests:



ISSN No:-2456-2165

Antibiotic Sensitivity Test

To test the susceptibility of *Staphylococcus Species* towards antibiotics.

1. Mueller Hinton Agar media was prepared for different bacterial cultures.

2. Prepared discs of Hexa G plus 1 and Hexa G plus 5 were used.

3. Mueller Hinton Plates were inoculated with different bacterial culture and prepared discs were placed on the plates and incubated for 24 hours.

4. Zone of inhibition were observed and studied.

# **RESULTS AND DISCUSSION**

In the result, two species of *Staphylococcus Species* were isolated. They are-

- Staphylococcus aureus and
- Staphylococcus epidermidis

Study on their enzyme activity through biochemical test and susceptibility towards different antibiotics were also studied over *Staphylococcus Species*.



Subculturing plates of *staphylococcus species*. pure culture of *staphylococcous epidermidis*.



Plate showing cultures of two different species pure culture of *staphylococcus aureus* of *staphylococcus* from slum area in mannitol salt agar.



Culture plate of *Staphylococcus Epidermidis* pure cultures of *Staphylococcus Aureus and* From railways station sample *Staphylococcus Epidermidis* 



Pure culture plate of *Staphylococcus species* NAM media showing *Staphylococcus aureus* & *Staphylococcus epidrmidis*.

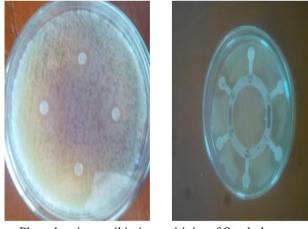


Plate showing antibiotic sensitivity of Staphylococcus.

ISSN No:-2456-2165

### DISCUSSION

S. Aureus and S. Epidermidis have emerged as an important cause of infections in patients with indwelling medical devices. Slime as a virulence factor of S. Epidermidis in medical implant/device related infections was documented in the past. It was also shown that comparatively higher numbers of slime producing isolates of S. epidermidis from the keratitic lesions were adherent to artificial surfaces as compared to the commensally isolates from the eye. Breakage of the skin barrier may lead to the transformation of S. aureus from a commensally colonizer to an invading pathogen, and its multitude of virulence factors enables it to adhere to and survive on and in the host cells. In most categories of hospitalized patients, S. aureus nasal carriage has been identified as a major risk factor for developing subsequent infections. Asserting that S. aureus infections are of endogenous origin is supported by studies revealing that isolates from nosocomial S. Aureu s infections were identical to nasal carrier isolates in 80% or more of the patients. In nonsurgical patients who were S. ureus nasal carriers, the risk of acquiring a nosocomial S. aureus bacteraemia was three times higher than in no carriers. To gain an understanding of S. aureus nasal carriage and the connection with subsequent infection, the S. aureus population structure needs to be defined. In our studies, we investigated the population structure of S. aureus in adult nasal carriers from a general population and among healthcare workers.

#### ACKNOWLEDGMENTS

Authors are thankful to Dr. D.K.Dubey, Principal of Sadhu Vaswani College,Bairagarh,Bhopal for providing lab facilities to carry out the above work.

## REFERENCES

- Aly R: Cutaneous microbiology. In: M Orkin, HI Maibach, MV Dahl, eds. Dermatology. Los Altos: Appleton & Lange, 1991.
- [2]. Feingold DS: Bacterial adherence, colonization, and pathogenicity. Arch Dermatol 122:161±163, 1986.
- [3]. Gao Z., Tseng C.H., Pei Z., Blaser M.J. Molecular analysis of human forearm superficial skin bacterial biota. Proc. Natl. Acad. Sci. 2007;104:2927–2932. [PMC free article] [PubMed].
- [4]. Goto H, Shimada K, Ikemoto H, Oguri T (2009). Study Group on Antimicrobial Susceptibility of Pathogens Isolated from Respiratory Infections. Antimicrobial susceptibility of pathogens isolated from more than 10,000 patients with infectious respiratory diseases: A 25- year longitudinal study. J. Infect. Chemother., 15: 347-360.

- [5]. Holland K.T., Cunliffe W.J., Roberts C.D. Acne vulgaris: An investigation into the number of anaerobic diphtheroids and members of the Micrococcaceae in normal and acne skin. Br. J. Dermatol. 1977;96:623– 626. [PubMed].
- [6]. Hugenholtz P., Pace N.R. Identifying microbial diversity in the natural environment: A molecular phylogenetic approach. Trends Biotechnol. 1996;14:190– 197. [PubMed] Pace N.R. A molecular view of microbial diversity and the biosphere. Science. 1997;276:734– 740.[PubMed].
- [7]. Mims C, Dockrell HM, Goering RV. (2004) Medical Microbiology. 3 editions. Elsevier Mosby, Edinburgh, United Kingdom: 2004; 585-586.Sandilands A., Terron-Kwiatkowski A., Hull P.R., O'Regan G.M., Clayton T.H., Watson R.M., Carrick T., Evans A.T., Liao H., Zhao Y., et al. Comprehensive analysis of the gene encoding filaggrin uncovers prevalent and rare mutations in ichthyosis vulgaris and atopic eczema. Nat. Genet. 2007;39:650–654.[PubMed].
- [8]. Ryan J. Kenneth, Ray C. George. (2003) An introduction to infectious diseases (2003). Sherris Medical Microbiology 4<sup>th</sup> Ed.
- [9]. Segre J.A. Epidermal barrier formation and recovery in skin disorders. J. Clin. Invest. 2006;116:1150– 1158. [PMC free article] [PubMed].
- [10]. Thomsen R.J., Stranieri A., Knutson D., Strauss J.S. Topical clindamycin treatment of acne. Clinical, surface lipid composition, and quantitative surface microbiology response. Arch. Dermatol. 1980;116:1031– 1034. [PubMed].
- [11]. Till A.E., Goulden V., Cunliffe W.J., Holland K.T. The cutaneous microflora of adolescent, persistent and lateonset acne patients does not differ. Br. J. Dermatol. 2000;142:885–892. [PubMed].
- [12]. Waldvogel F. D. (1990) Staphylococcus aureus (including toxic shock syndrome) In: Mandell GL, Donglas RG, Bonnett JE (Eds.). Principles and practice of Infectious Disease, 3<sup>rd</sup> ed. Churchill Livingston, London. Pp 1489- 1510.